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## A Tutorial for Chemists: Using Mnova to Process, Analyze and Report 1D and 2D NMR on Your Desktop

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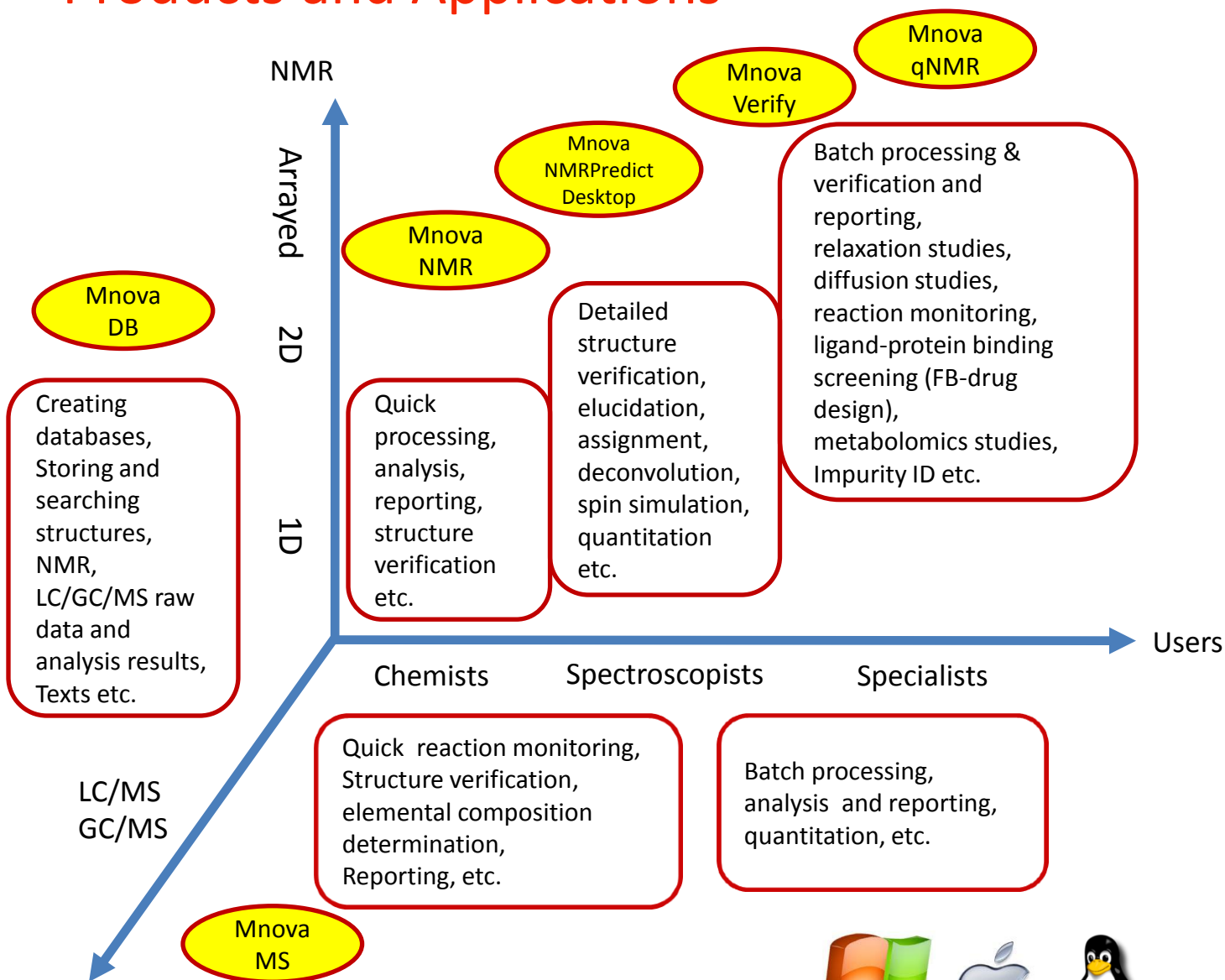
# Outline

- M** Overview of Mestrelab and Mnova
- M** Open and process 1D and 2D NMR data
- M** Multiplet Analysis for 1D H-1 NMR
- M** Assign 1D peaks to a structure
- M** Assign 1D and 2D spectra
- M** Report analysis results
- M** Basic handling of multiple spectra

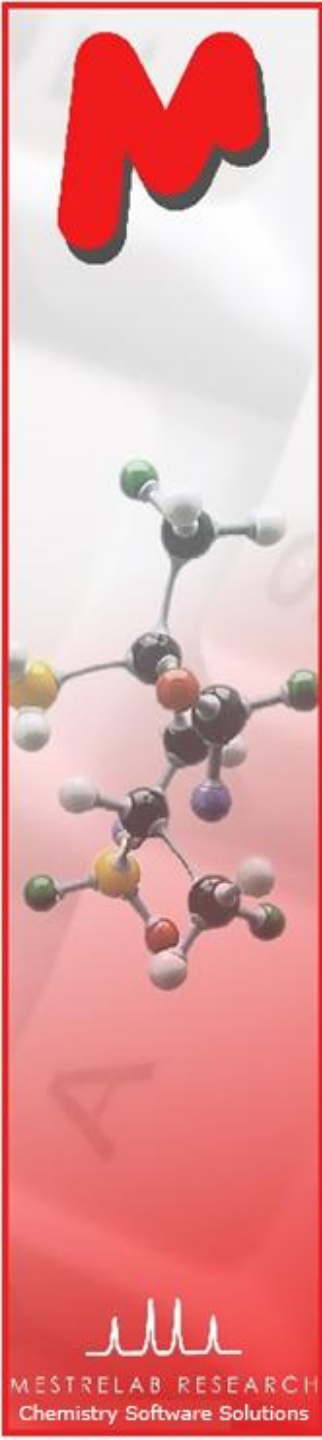
*Note: Only the basic features of Mnova NMR (and NMRPredict Desktop) are covered in this tutorial. For information about the advanced features of Mnova NMR, see <http://mestrelab.com/software/mnova-nmr/>. For information about other plugins in Mnova, such as MS, DB, Verify, qNMR, and Screen, please see <http://mestrelab.com/software/> or write to [chen.peng@mestrelab.com](mailto:chen.peng@mestrelab.com).*



# Products and Applications



Mnova is compatible with Mac, Windows and Linux





# Mestrelab Research

- M** 1996: A research project in University of Santiago de Compostela, Spain, developed free **MestReC** software for NMR processing
- M** 2004: **Mestrelab Research** incorporated in Santiago de Compostela
- M** 2004: New **MestreNova (Mnova)** platform and **NMR** plugin released
- M** 2006: **NMRPredict Desktop** plugin released with Modgraph
- M** 2009: **LC/GC/MS** plugin released with Sierra Analytics
- M** 2009: Global Spectral Deconvolution (**GSD**) algorithm released with ExtraByte
- M** 2011: **DB** plugin for Database Management
- M** 2012: **Verify** plugin for auto structure verification and peak assignment
- M** 2012: **qNMR** plugin for quantitative NMR analysis - to be released
- M** 2013: **Screen** plugin for high-throughput ligand-protein binding analysis - to be released
- M** An **R&D company** with >20 people and >80,000 registered users

[www.mestrelab.com](http://www.mestrelab.com)



# Before you start

- M Make sure Mnova is properly installed and licensed on your computer.
- M If you don't have license, you can download, install and free trial for 45 days\*: <http://mestrelab.com/software/mnova-suite/download/>
- M To verify your licensing status, choose **Help > License Manager**

Host ID (unique for this computer)

State	Plug-in	Issued By	Licensed To	Type	Issue Date	Days to Expire	Update Days	Valid Days	Path
1 ?	ASV Batch	Mestrelab Res...	Chen Peng	single	Wed May 9 2012	1365	1365	N/A	C:/Users/Chen Peng/AppData... {57f176a7-2b6d...
2 ✓	DbPlugin	Mestrelab Res...	Chen Peng	single	Fri Sep 24 2010	Never	1137	N/A	C:/Users/Chen Peng/AppData... {f04ec48f-5904...
3 ✓	Mass	Mestrelab Res...	Chen Peng	single	Fri Sep 24 2010	Never	1137	N/A	C:/Users/Chen Peng/AppData... {d1f3481f-2037...
4 ?	MSpin	Mestrelab Res...	Chen Peng	single	Fri Sep 24 2010	Never	1137	N/A	C:/Users/Chen Peng/AppData... {967c36fe-e527...
5 ✓	NMR	Mestrelab Res...	Chen Peng	single	Fri Sep 24 2010	Never	1137	N/A	C:/Users/Chen Peng/AppData... {27fece3a-5d55...
6 ✓	Modgraph NMRPredict Desktop	Mestrelab Res...	Chen Peng	single	Fri Sep 24 2010	Never	1137	N/A	C:/Users/Chen Peng/AppData... {2dce56a7-62c4...
7 ?	Reaction Monitoring	Mestrelab Res...	Chen Peng	single	Wed May 9 2012	1365	1365	N/A	C:/Users/Chen Peng/AppData... {56413c8d-f9a3...
8 ✓	Mnova Screen	Mestrelab Res...	Chen Peng	single	Wed May 9 2012	1365	1365	N/A	C:/Users/Chen Peng/AppData... {522b110d-fafc...
9 ✓	Automatic Structure Verification	Mestrelab Research S.L.	Dr. Chen Peng	single	Fri Mar 30 2012	594	594	N/A	C:/Users/Chen Peng/AppData/Roaming/Mes... {d6e0a3e-9627...

Make sure you have a positive number or "Never" (for perpetual license) here

Days before your Update and Support package expires\*\*

Location of the license files

\*There is no difference between the free trial version and the released version of Mnova.

\*\*If your Update and Support package has expired, you can only run the versions of Mnova that were released before the expiration date. Find previous versions of Mnova at <http://mestrelab.com/software/mnova-suite/download/versions/>.



## Sample data sets

- M** A set of 1D  $^1\text{H}$ ,  $^{13}\text{C}$ , 2D HSQC, and LC/MS\* raw data of quinine are included when Mnova is installed. So is the .mol file. You can find them in a folder similar to the following:  
C:\Program Files (x86)\Mestrelab Research S.L\MestReNova\examples\datasets
- M** These data are used in many of the examples shown in this tutorial.
- M** When you use your own data, make sure you copy all the files in the folder of an experiment.

*\*You need the Mnova MS license to open and analyze LC/MS data. See <http://mestrelab.com/software/mnova-ms/> for details.*

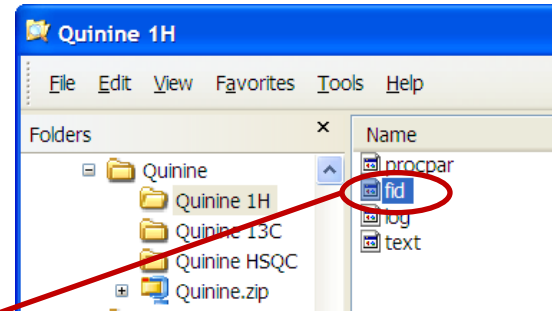
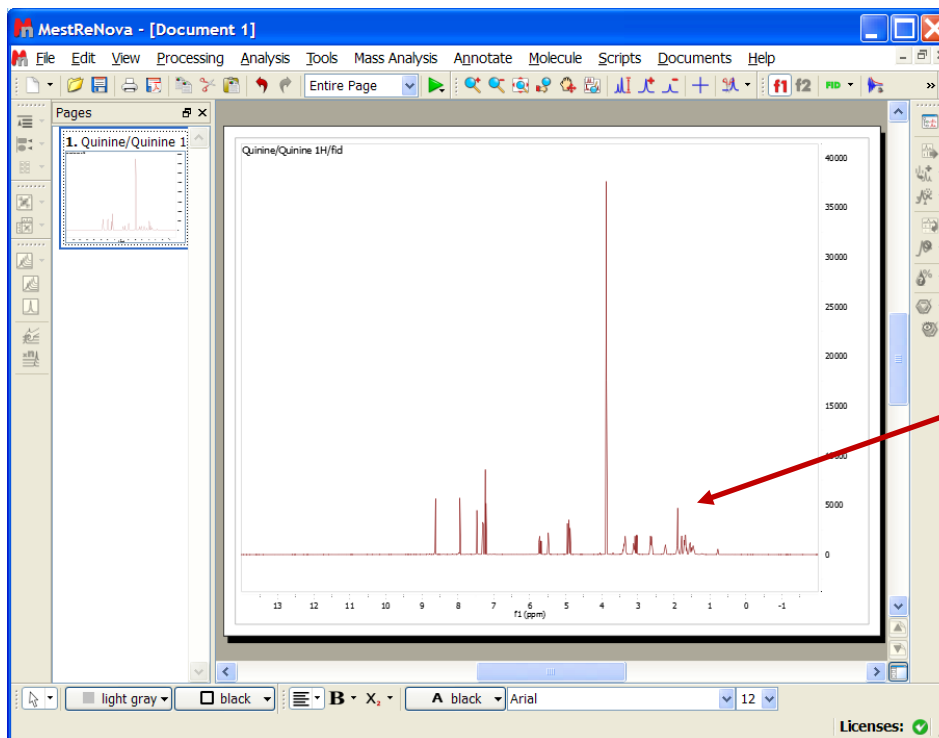




# M

## To open and transform your NMR data

- M Choose **File | Open** to open the **fid** (or **ser**) file from the raw data
- M Or drag an **fid** file from a file browser to Mnova \*
- M Mnova automatically transforms the raw file into frequency domain (including *Windowing function, Fourier transform, phase correction etc*) \*\*



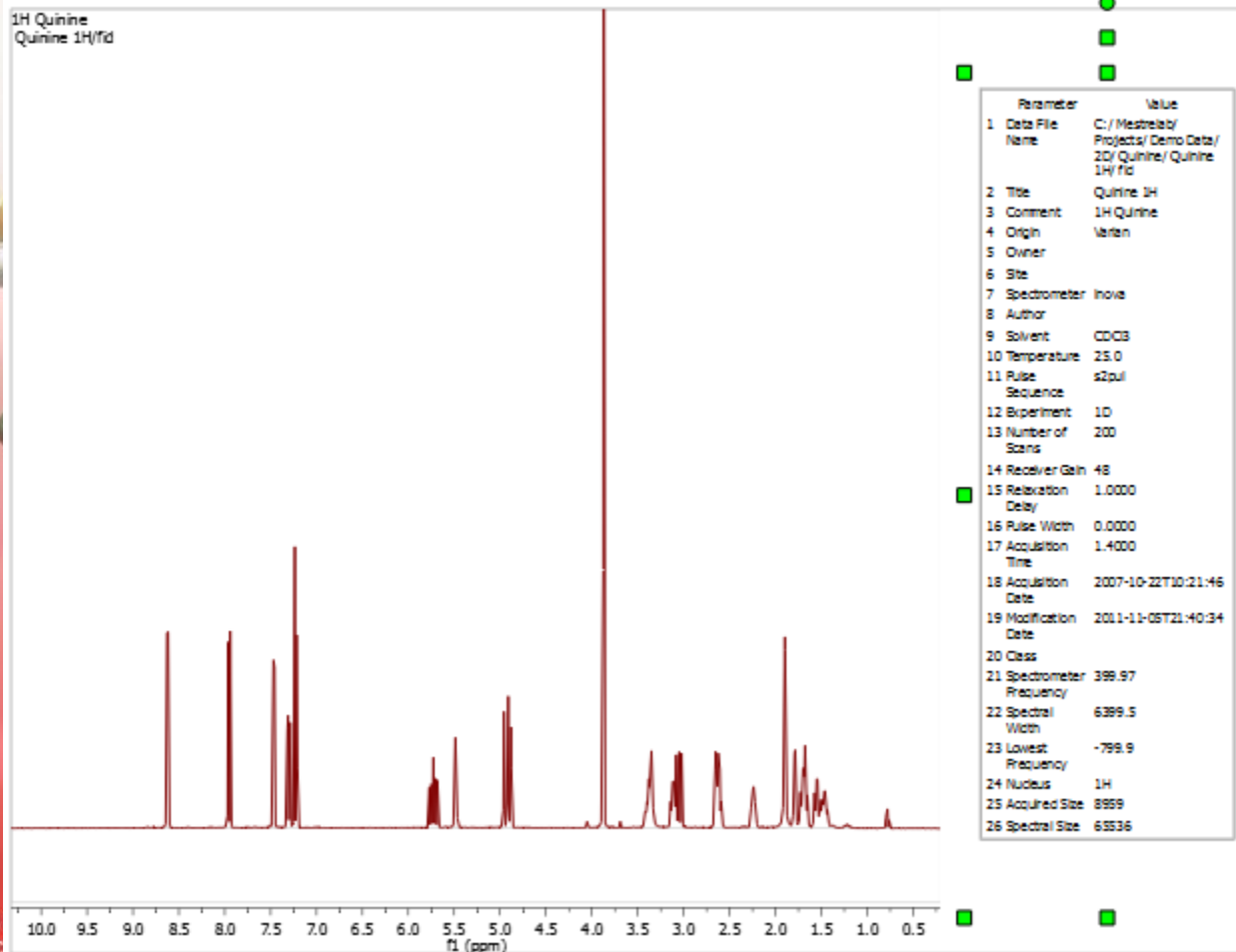
Drag & drop

- \*You can drag **multiple folders** that contain **fid** (or **ser**) files to Mnova to open multiple spectra simultaneously.
- \*\*Parameters from the raw data are used for processing. You can control the importing of some parameters (zero filling, phasing, baseline correction etc) by choosing **Edit > Preferences > NMR > Import**. You can view or change the processing parameters by choosing **Processing | Processing Parameters**.



# To see the parameters

- Choose **View | Tables | Parameters** to view the acquisition and processing parameters
- Click **Report** to report the parameters as a text box on the spectrum. Resize the text box and spectrum to make a better layout



Parameters


Report Copy Setup Customize

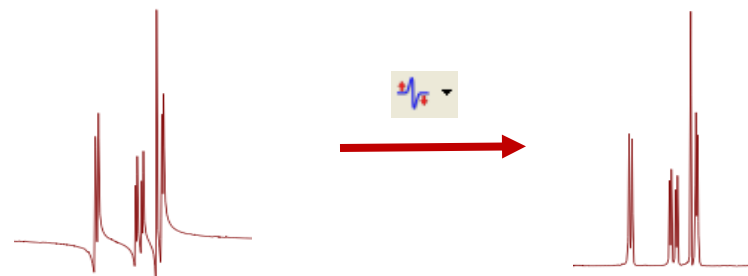
Parameter	Value
1 Data File Name	C:/Mestrelab/Projects/Demo Data/2D/Quinine/Quinine 1H/fid
2 Title	Quinine 1H
3 Comment	1H Quinine
4 Origin	Varian
5 Owner	
6 Site	
7 Spectrometer	inova
8 Author	
9 Solvent	CDCl3
10 Temperature	25.0
11 Pulse Sequence	s2pul
12 Experiment	1D
13 Number of Scans	200
14 Receiver Gain	48
15 Relaxation Delay	1.0000


Use the green handles to move, rotate and resize the text box

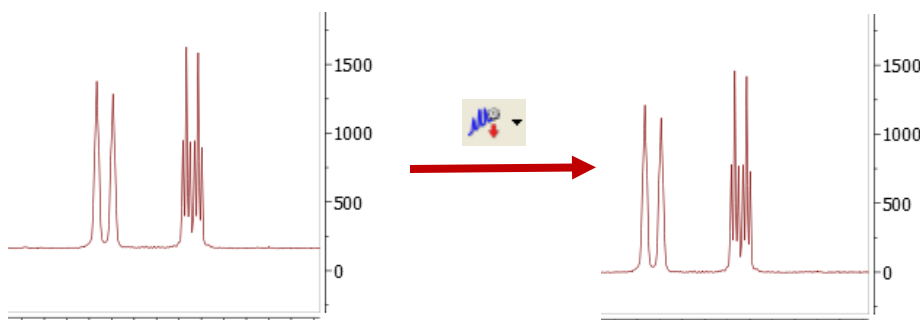



# To correct phase, baseline & reference

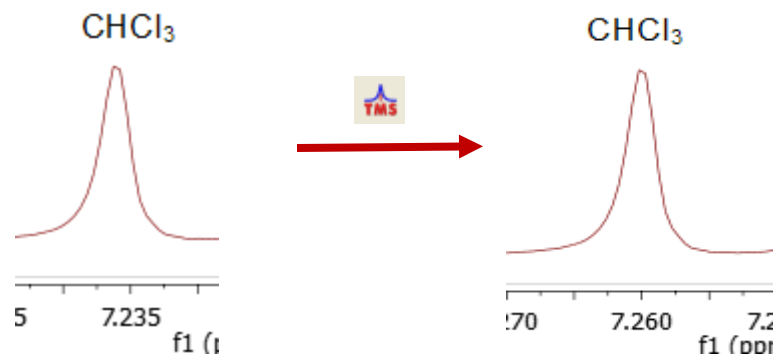
- Click  for **phase correction** if peaks are not symmetric\*



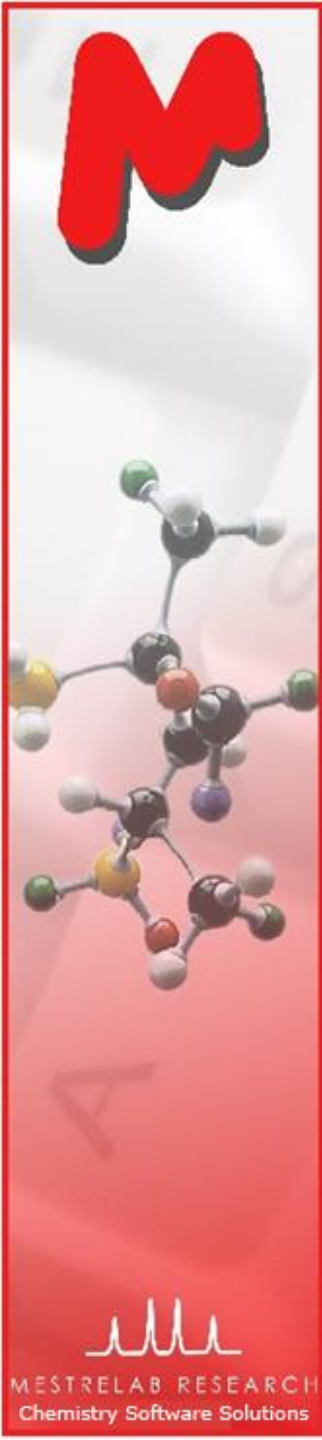
- Click  for **baseline correction** if baseline is not zero \*



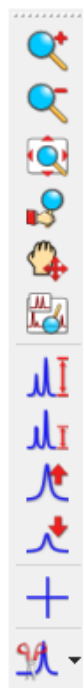
- Click  to calibrate the **chemical shift reference** if the solvent or TMS peak is not at the right ppm



\*Click the arrow next to the tool icon for options, such as manual phasing and manual baseline correction. See **Help > Contents > Processing Basics** for more details



# To visualize your spectrum



Zoom in/Zoom out (or press Z) \*

Zoom out

Full spectrum (or press F)

Manual Zoom in to defined ppm range

Pan spectrum (or press P)\*\*

Expansion – click&drag to draw an inset (or press E)

Fit to Highest Intensity (or press H)

Fit to highest compound peak

Increase Intensity (or rotate mouse wheel)

Decrease Intensity (or rotate mouse wheel)

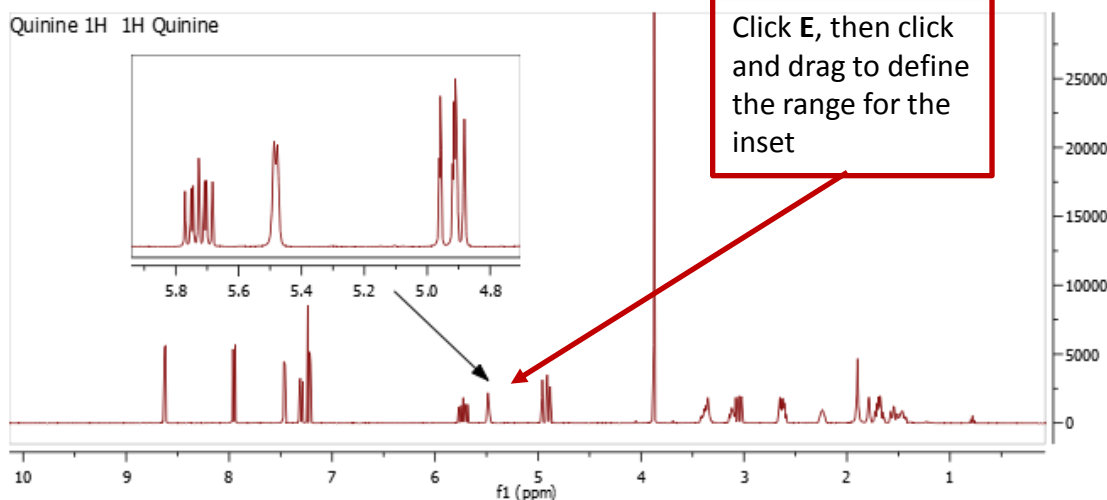
Crosshair Cursor (or press C) for measuring  $J$ -couplings

Cut (or press X) to hide parts of the spectrum

*\*Press Z several times to toggle between horizontal/vertical/box zoom*

*\*\* Press P several times to toggle between free/horizontal/vertical panning*

Quinine 1H 1H Quinine

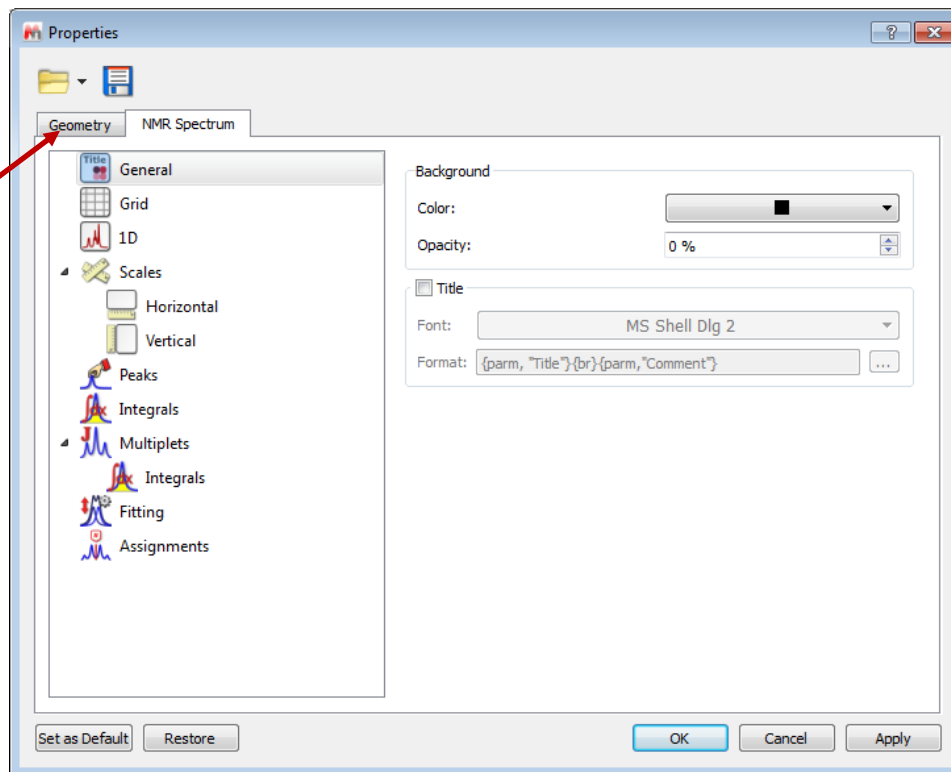


Click E, then click and drag to define the range for the inset

# To change the display properties

- M** Right click on a spectrum and choose **Properties** from the context menu
- M** A lot of display properties can be customized
- M** You can click **Set as Default** to save the settings for spectra opened in the future
- M** You can save the settings for other users using the **Save Properties** and **Load Properties** tools

**Tip:** Use options in the **Geometry** Tab to change the size of a spectrum precisely

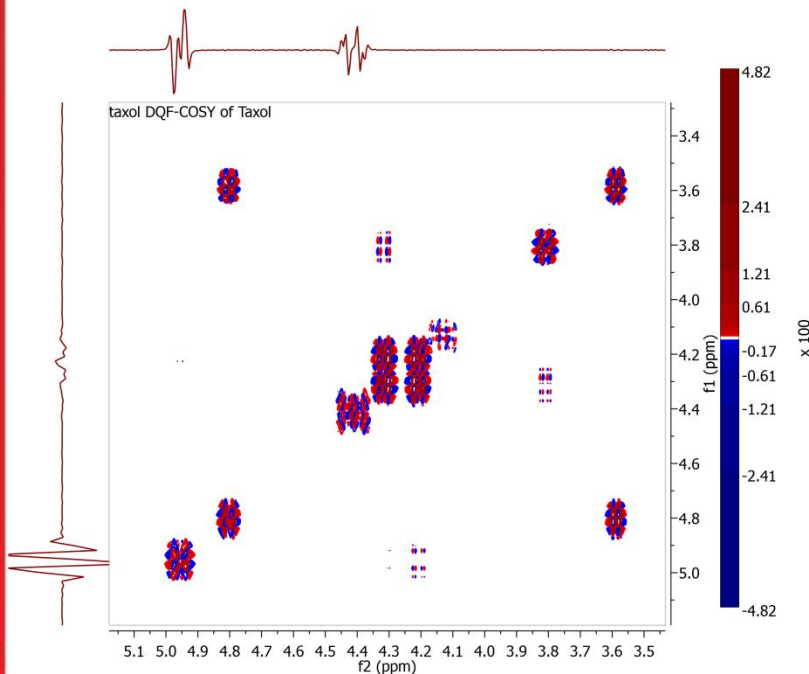
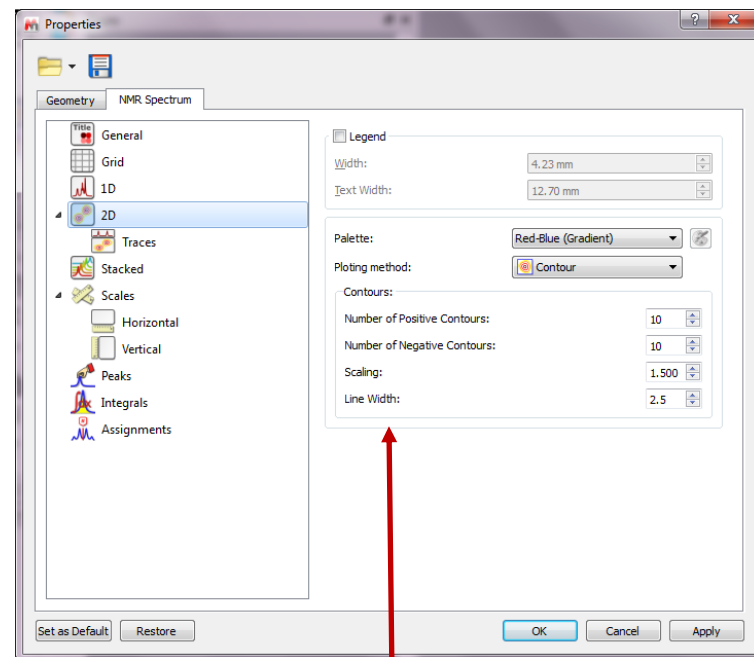
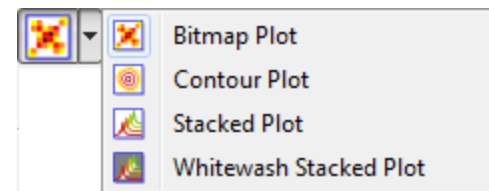


# M

## To display 2D spectra in the way you want

- Use the **Plot Mode** tools to change to bitmap or contour display etc.
- You can also change other display properties by right-clicking on the spectrum and then choose **Properties**:



- Legend
- Color Palette: You can define your own
- Contours: numbers and scaling, and line width
- Traces: method and space

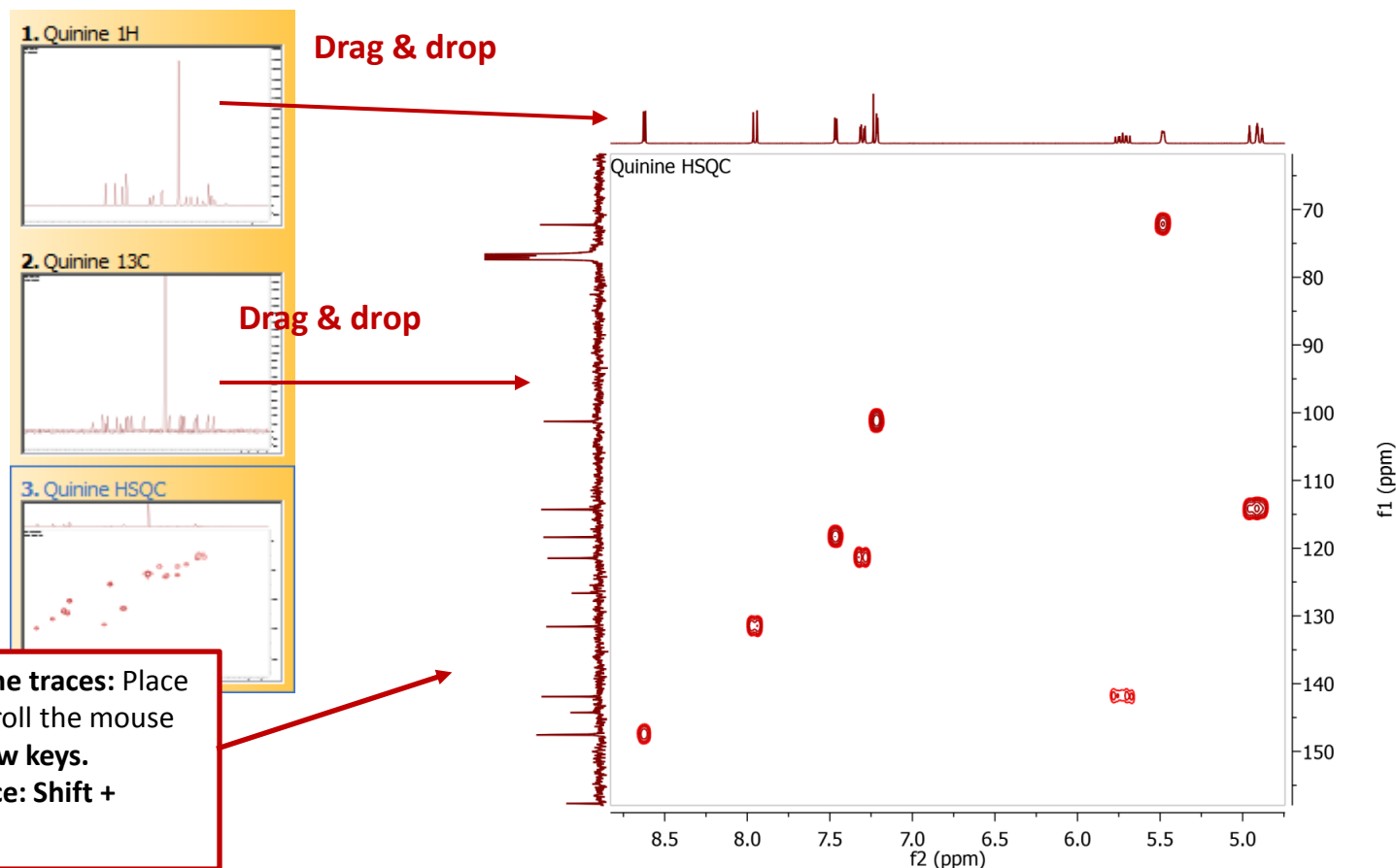
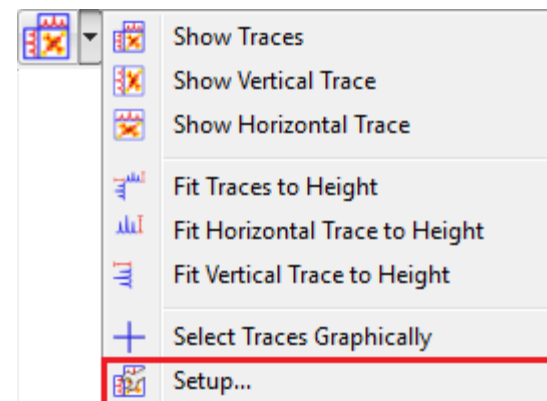


**Tip:** You can set a line width for 2D contours independent of that for 1D curves since 7.1.1

# M

## To attach 1D to 2D spectra


- 1 Open 1D and 2D spectra in the same document (They are shown as separate pages)
- 2 Display the 2D spectrum, click the **Traces** tool options  and choose **Setup...**
- 3 Choose a 1D in the Available 1D Spectrum, click  to attach it to that axis




**To change the Y intensity of the traces:** Place the cursor on the trace and scroll the mouse wheel, or click **Ctrl+Shift+arrow keys**.  
**To move the baseline of a trace:** Shift + mouse wheel.

# To analyze and report multiplets of H-1 NMR

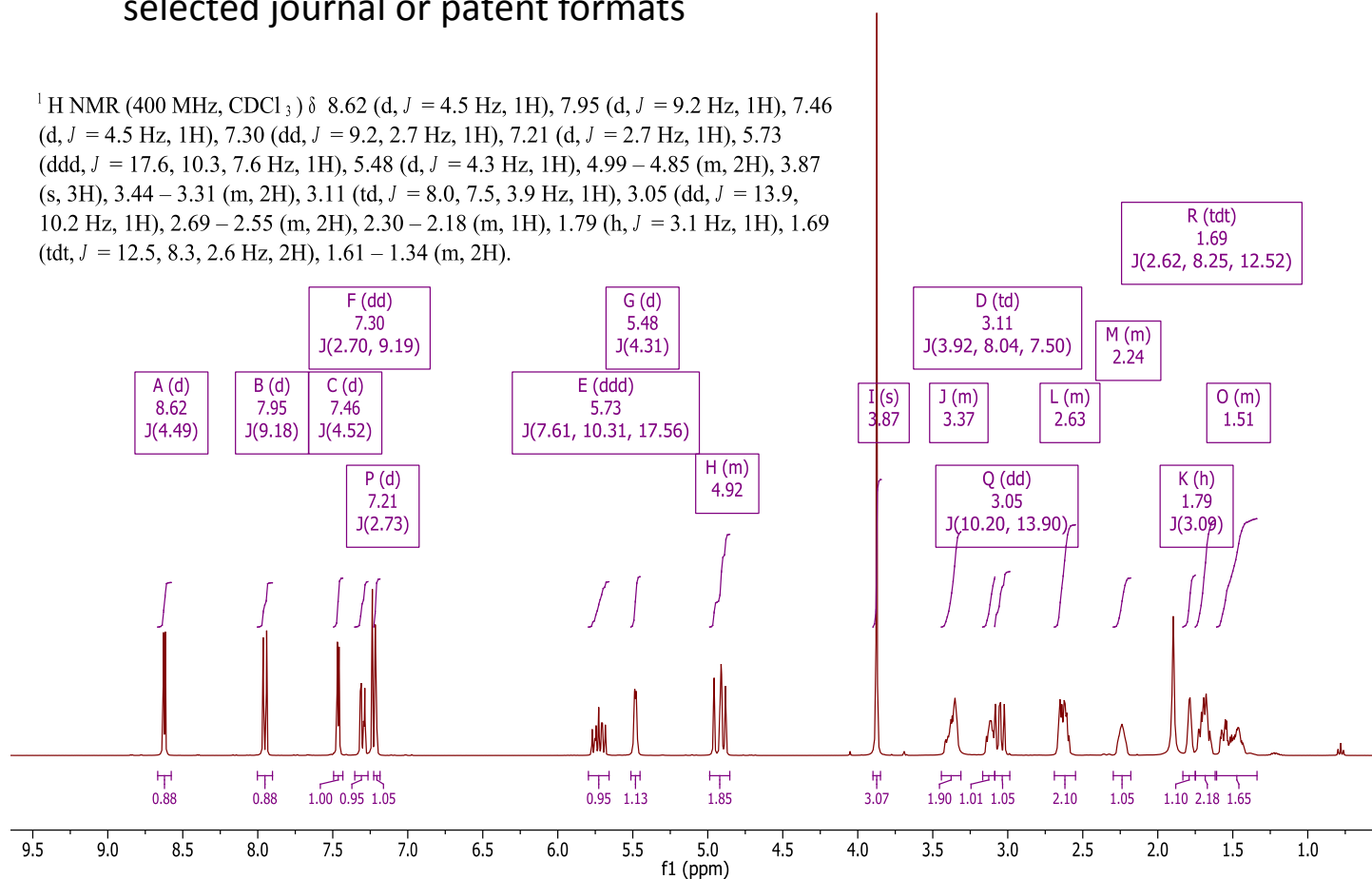
M Mnova provides two approaches to **multiplet analysis**:

M  **Fully automatic**: peak picking, integration and multiplet analysis *all done by one click*, with peaks deconvolved using GSD\* and types classified

M  **Manual**: click-and-drag to pick each multiplet interactively

M In either case you can **refine** the results interactively, and **report** them in selected journal or patent formats



<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.62 (d, *J* = 4.5 Hz, 1H), 7.95 (d, *J* = 9.2 Hz, 1H), 7.46 (d, *J* = 4.5 Hz, 1H), 7.30 (dd, *J* = 9.2, 2.7 Hz, 1H), 7.21 (d, *J* = 2.7 Hz, 1H), 5.73 (ddd, *J* = 17.6, 10.3, 7.6 Hz, 1H), 5.48 (d, *J* = 4.3 Hz, 1H), 4.99 – 4.85 (m, 2H), 3.87 (s, 3H), 3.44 – 3.31 (m, 2H), 3.11 (td, *J* = 8.0, 7.5, 3.9 Hz, 1H), 3.05 (dd, *J* = 13.9, 10.2 Hz, 1H), 2.69 – 2.55 (m, 2H), 2.30 – 2.18 (m, 1H), 1.79 (h, *J* = 3.1 Hz, 1H), 1.69 (td, *J* = 12.5, 8.3, 2.6 Hz, 2H), 1.61 – 1.34 (m, 2H).



\*GSD (Global Spectral Deconvolution): See Help > Contents > Analysis tools > Peak Picking > GSD for details



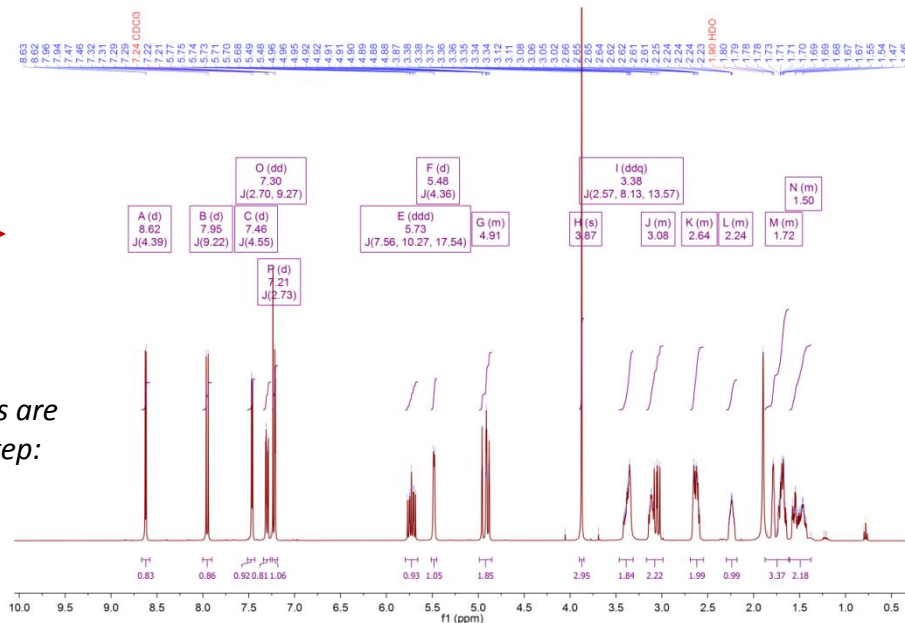
# Alert to users of Version 6 or older: Adopt the new workflows for more efficient multiplet analysis

- There have been major changes to the peak picking, integration and multiplet analysis since Version 7, including GSD, auto peak classification, and more intelligent multiplet analysis.
- If your goal is to extract multiplet information (chemical shifts, integrals and  $J$ -couplings), use the **Automatic Multiplet Analysis tool**  or the **Manual Multiplet Analysis tool**  to extract such info *directly*.
- Do NOT integrate peaks prior to it.** If you did manual or auto integration prior to multiplet analysis, such integrals will be replaced by the multiplet integrals. Integrals from either operations are independent, and may be different due to different integration mechanisms and options.



You are recommended to use the **Automatic Multiplet Analysis** or **Manual Multiplet Analysis** directly **without** a priori peak picking or integration

*Pick picking and classification, integration, and multiplet analysis are all done in one step:*






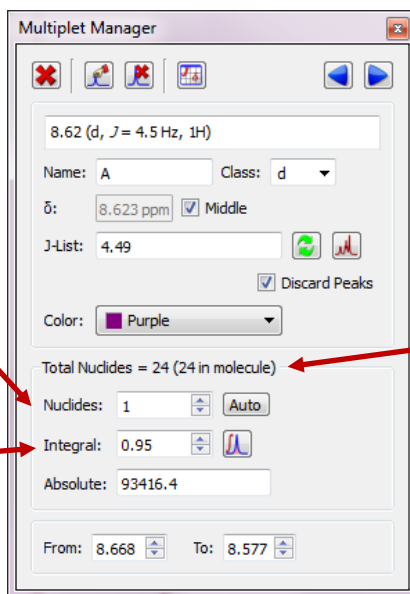
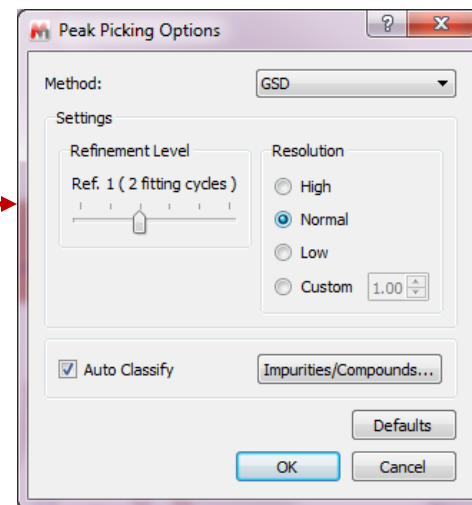
# M

## Fully automatic multiplet analysis



Click  to do automatic multiplet analysis. By default, it does the following:

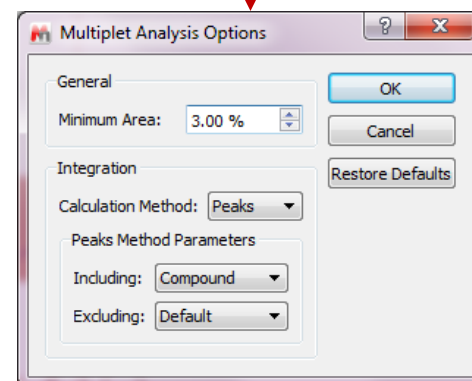
- Picks peaks using GSD (if no peaks were picked) and classify their types (compound, solvent, impurity peaks etc.). Note these are controlled by the Peak Picking options
- Groups the picked peaks into multiplets and fits them to  $J$ -coupling patterns, and calculates their integrals (depending on the Multiplet Analysis options). Note these are controlled by the Multiplet Analysis Options
- Estimates the total number of nuclides (NN) and normalizes the integrals for each multiplet



The number of nuclides (NN) of the multiplet


Normalized integral of the multiplet.

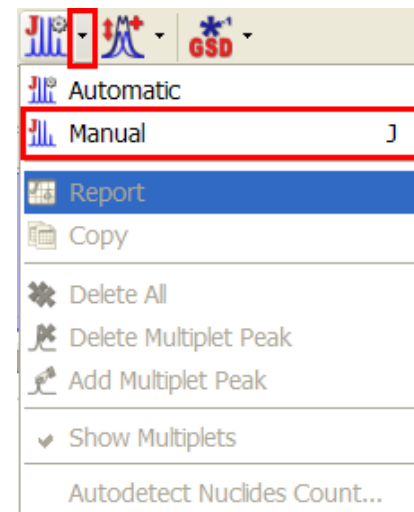
Total # of nuclides from all the multiplets and the # of protons in the molecule (if present)



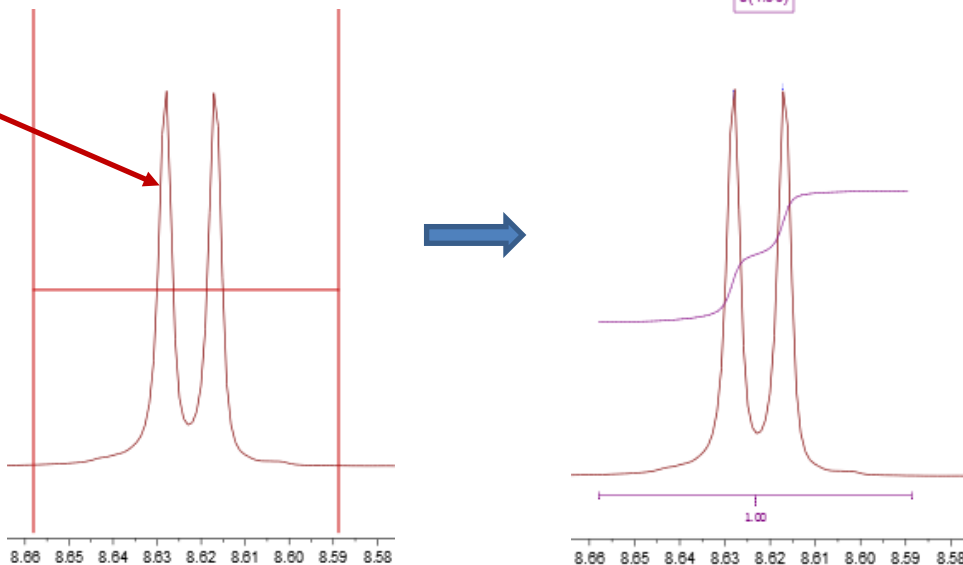


# To pick multiplets manually

- Manual Multiplet Analysis  allows you to have more control of the multiplet analysis (J is the shortcut key)
- You zoom into each multiplet, click and drag to define the following:
  - Peak picking threshold
  - Integration region\*
- Mnova picks the peaks in the region, fits them to a  $J$ -coupling pattern and defines the multiplet in the same way as in automatic multiplet analysis



Click and drag to define the **integration region** and **peak picking threshold** and a doublet will be picked



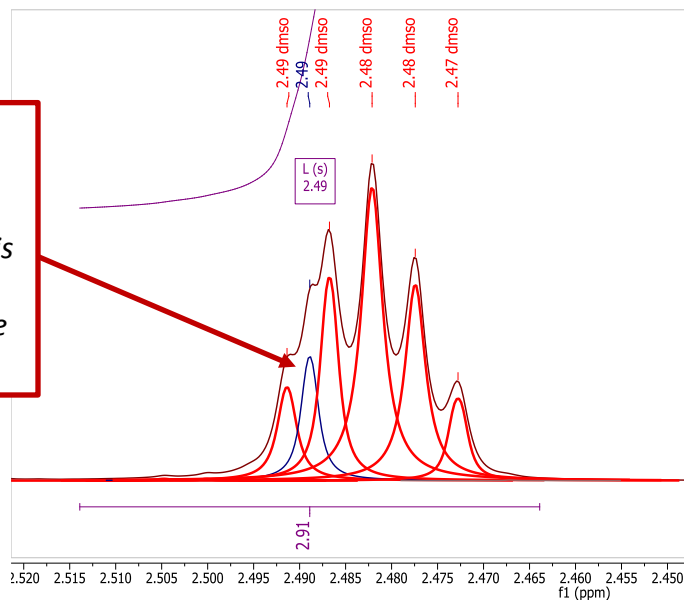
*Tip: To turn on the integral curves, right click and select Properties, go to Multiplets > Integrals.*

*\* If Peaks is used as the Integration Method, the area of a GSD peak will be included in the integral as long as the peak top falls within the region.*

# To manually refine the auto multiplet analysis results

- After the auto multiplet analysis, you are advised to use the Multiplet Manager to verify and correct the results if needed.
- The most important things to verify:
  - Are the solvent peaks properly identified, and impurity peaks properly excluded?
  - Are the integrals properly normalized and the numbers of nuclides correct?
  - Are the details of each multiplet correct?
- Mnova provides a set of tools for you to verify and correct such results interactively. The use of such tools are exemplified in the following slides.

*Tip: GSD-based multiplet analysis usually gives reliable deconvolution of overlapping peaks, as in this case. However, you are always advised to verify the results manually.*

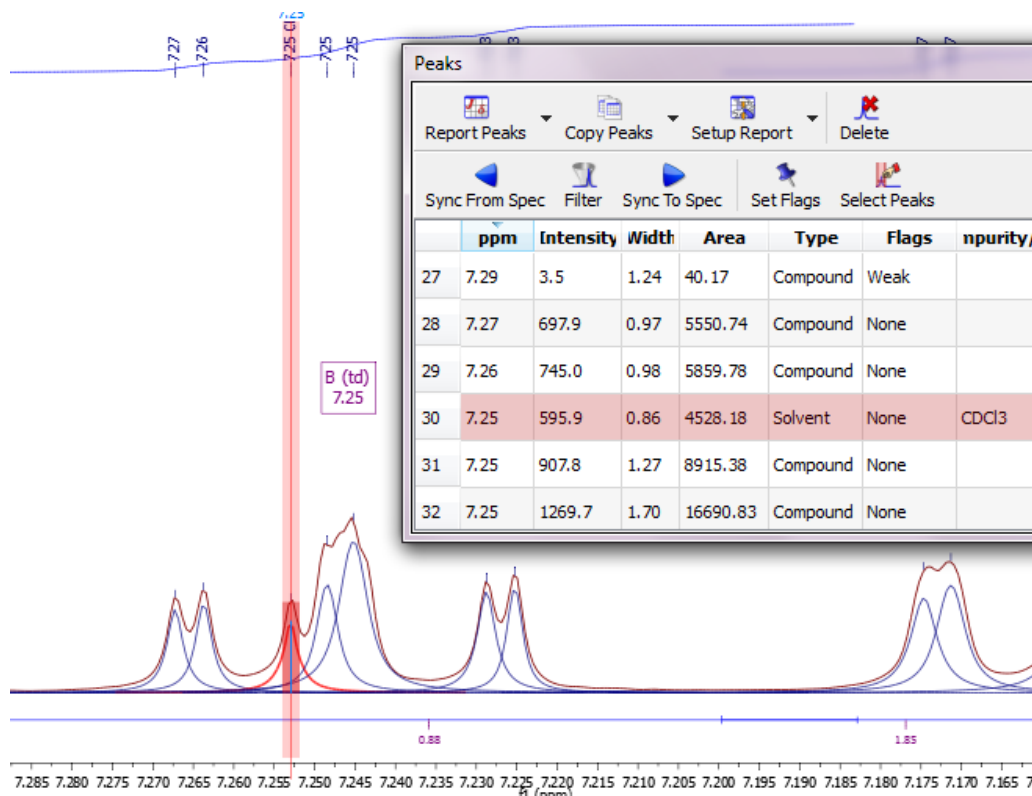



The screenshot shows the 'Multiplet Manager' window in Mnova. The main display shows a peak at 2.49 ppm (s, 3H). The 'Name' is 'L' and the 'Class' is 's'. The chemical shift  $\delta$  is 2.489 ppm, with the 'Middle' checkbox checked. The 'J-List' is empty, and the 'Discard Peaks' checkbox is checked. The 'Color' is set to 'Purple'. Below this, the 'Total Nuclides' is 19, the 'Nuclides' is 3 (with an 'Auto' button), the 'Integral' is 2.91, and the 'Absolute' value is 17844. At the bottom, the 'From' and 'To' values are 2.514 and 2.464, respectively.



# If solvent peaks are not properly recognized

- M Mnova has a sophisticated method for solvent peak recognition, though it may not always work right.
- M To view the peak types, turn on the peak curves to see the different colors, or click on any peak to open the **Peaks Table**:
- M To change the type of a peak, right click it from the spectrum, and choose **Edit Peak Type**.
- M You can also choose multiple peaks from the Peaks Table and change their types together

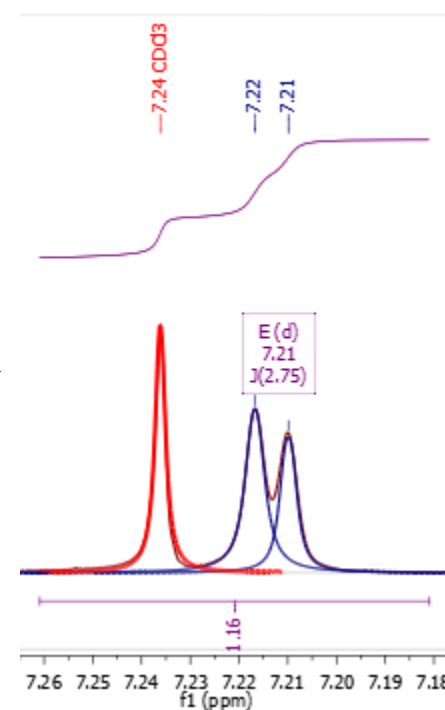
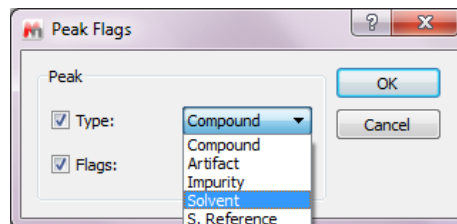
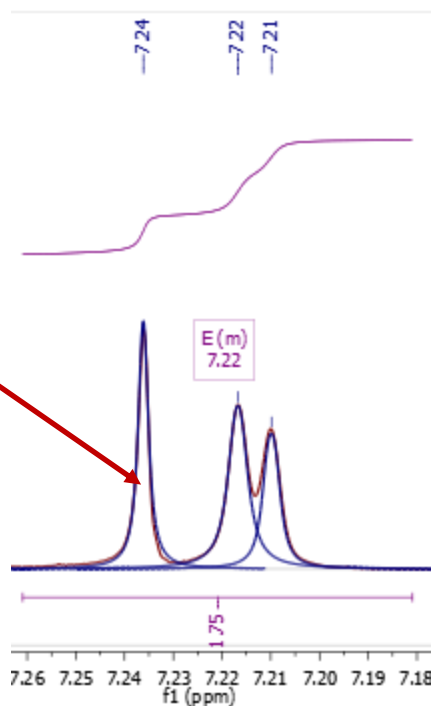


Tip: To display the GSD peak curves (and sum or residuals), expand the Peak Picking Tool menu  , and check Show Peak Curves or other options. Use the Properties dialog for more options.

## If solvent peaks are not properly recognized (2)

- If a wrong solvent peak affects only one multiplet, right click on that peak, choose Edit Peak Type, and change it to solvent/compound. This peak will be automatically excluded from/included to the multiplet:

Right click here and choose Edit Peak Type. Change its Type to Solvent



Note: The multiplet integral curve does not change to reflect the actual integral of the doublet. You can click on the left end of the multiplet bar and move it to the right to exclude the CDCl<sub>3</sub> peak from the integration curve. Note this does not change the integration value of the doublet.

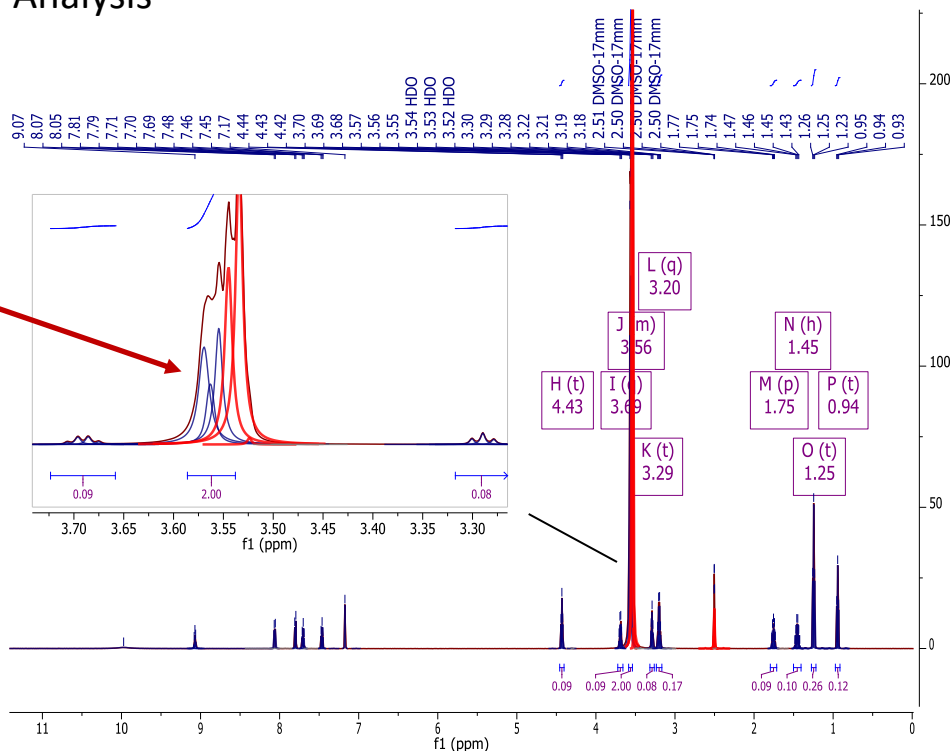




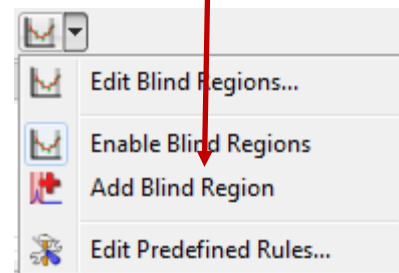
## If solvent peaks are not properly recognized (3)

- If a missed strong solvent peak affects the overall multiplet analysis results, try the following and redo the auto multiplet analysis:
  - Use the Edit Blind Regions tool to exclude the solvent peak(s).
  - In the Parameters Table, make sure the Solvent name is right.
  - Make sure Auto Classify is turned on in the Peak Picking Options
  - Or you can do manual peak picking first to exclude the solvent ones.
- Before redoing multiplet analysis, choose Analysis > Peak Picking > Delete All to remove all peaks (and hence the multiplets)
- If none of the above options works satisfactorily, use Manual Multiplet Analysis

As only part of the HDO peaks were marked as solvent peaks, auto multiplet analysis misses many small multiplets in the aromatic region



Use Add Blind Region to cover all the HDO peaks. Remove all peaks and do auto MA again to cover all multiplet peaks





# M

## Multiplet Manager

A (d)  
8.62  
J(4.39)

Double click on it to show the Multiplet Manager

- Double click on a multiplet label to open the Multiplet Manager. Use it to inspect and change the properties of the multiplets, including the normalization of the integrals,  $J$ -coupling patterns and constants etc.

Delete the current multiplet

Add/Delete multiplet peaks

Navigate to the Previous/Next multiplet

Properties of the current multiplet

The # of protons this multiplet corresponds to. Changing this number affects only the current multiplet

Use this tool to simulate the multiplet

Use this tool to measure J constant manually

Normalized integral of the multiplet. Changing it affects all multiplets\*

# of protons in the molecule (if present)

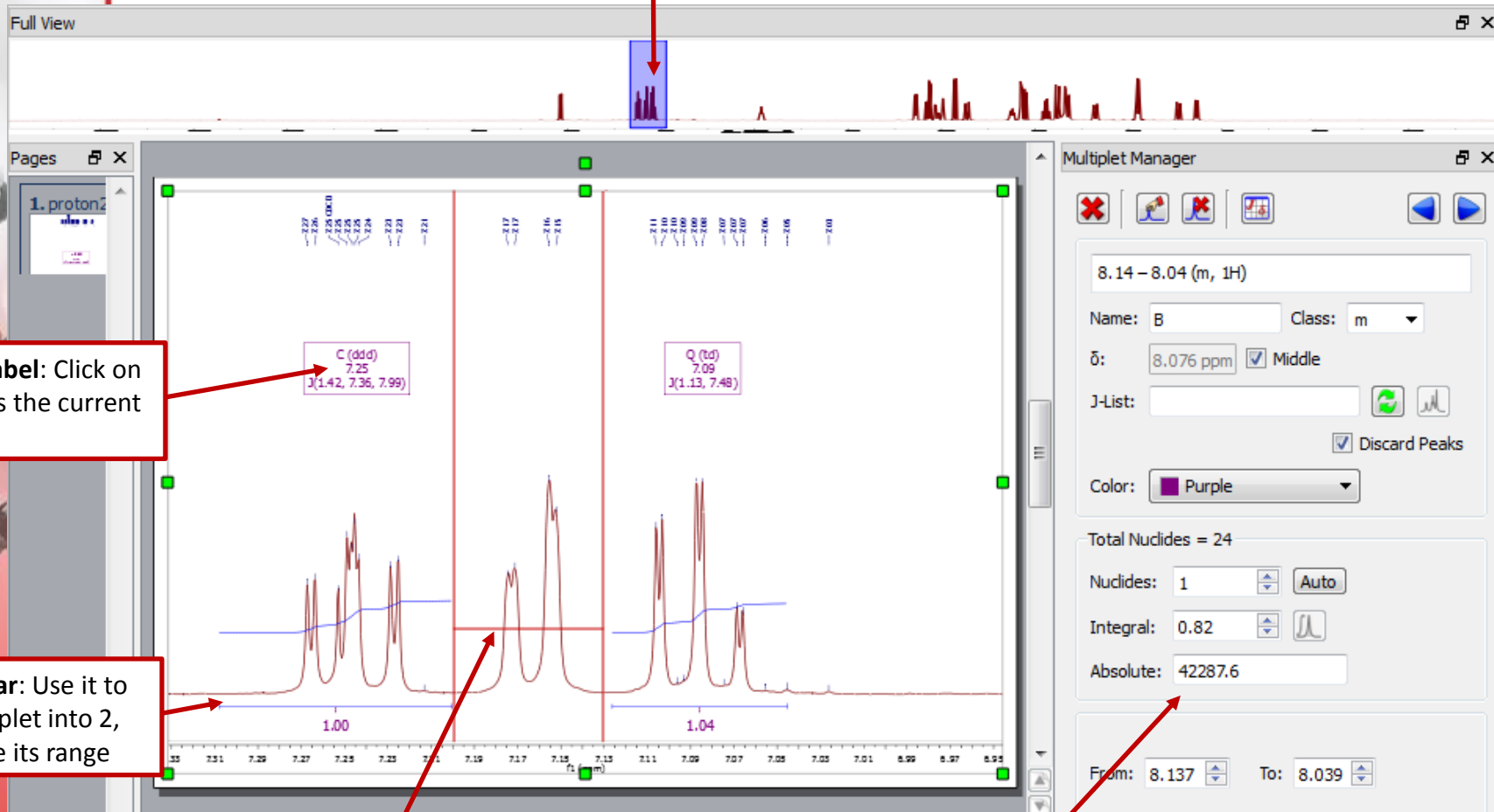
Integration region of the multiplet

Absolute integral of the multiplet

\*Note: the normalization here is **independent** of the normalization of integration using the Integral Manager.

# Handy tools for multiplet analysis

**Full View:** The whole spectrum and zoom-in area. Drag the blue box to move to other multiplets. (Choose **View | Full View** to open Full View)



**Multiplet label:** Click on it to set it as the current active one

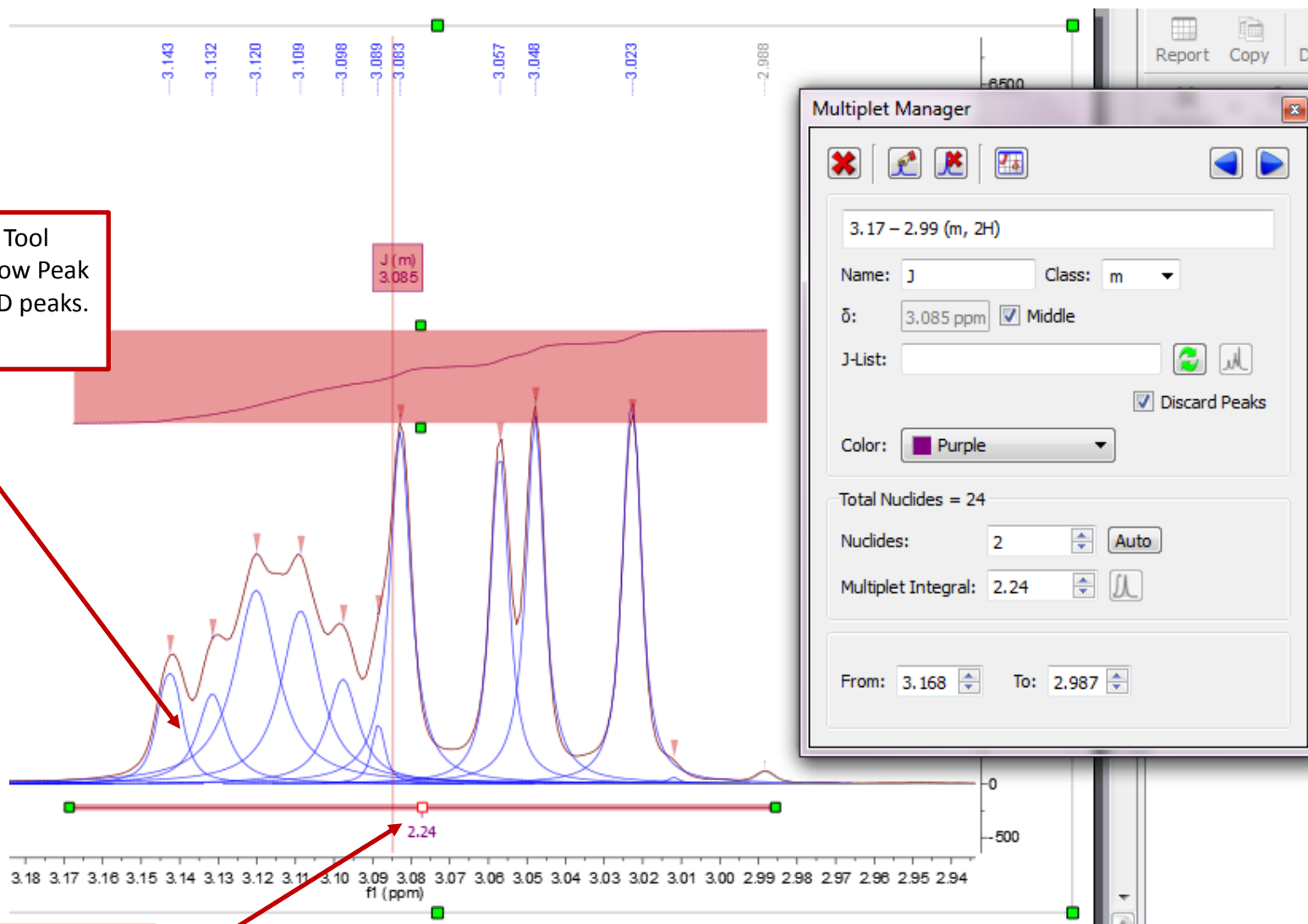
**Multiplet bar:** Use it to split a multiplet into 2, or to change its range


**Manual multiplet analysis:** Click J, then click and drag to define the range and peak picking threshold for a multiplet.

**Multiplet Manager** shows the properties of the current multiplet picked. (Double click on a multiplet label to open it)



# To split partially overlapping multiplets

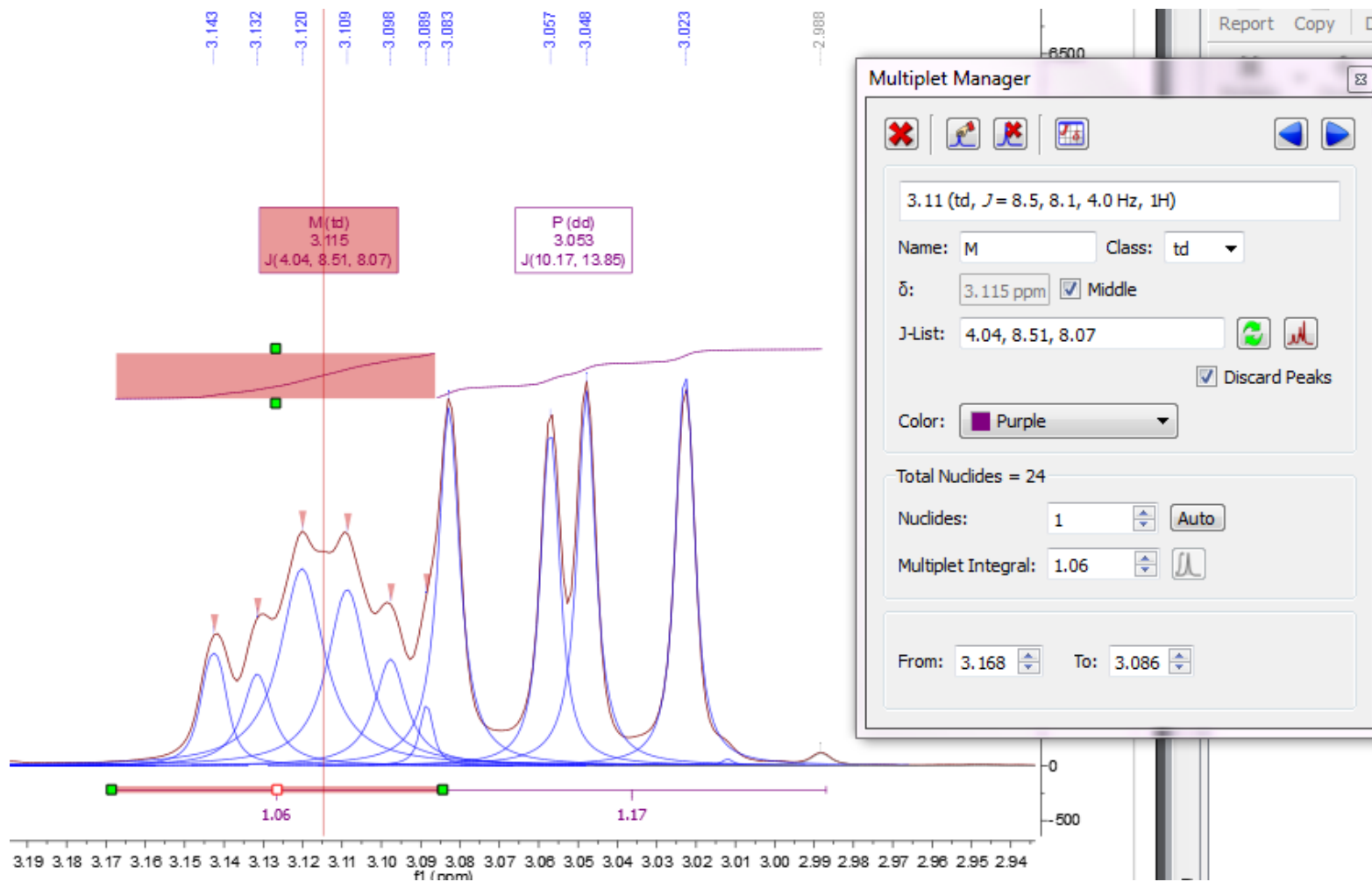


Expand the Peak Picking Tool menu  , check Show Peak Curves to display the GSD peaks.

Drag this red box to where you want to split the multiplet into two

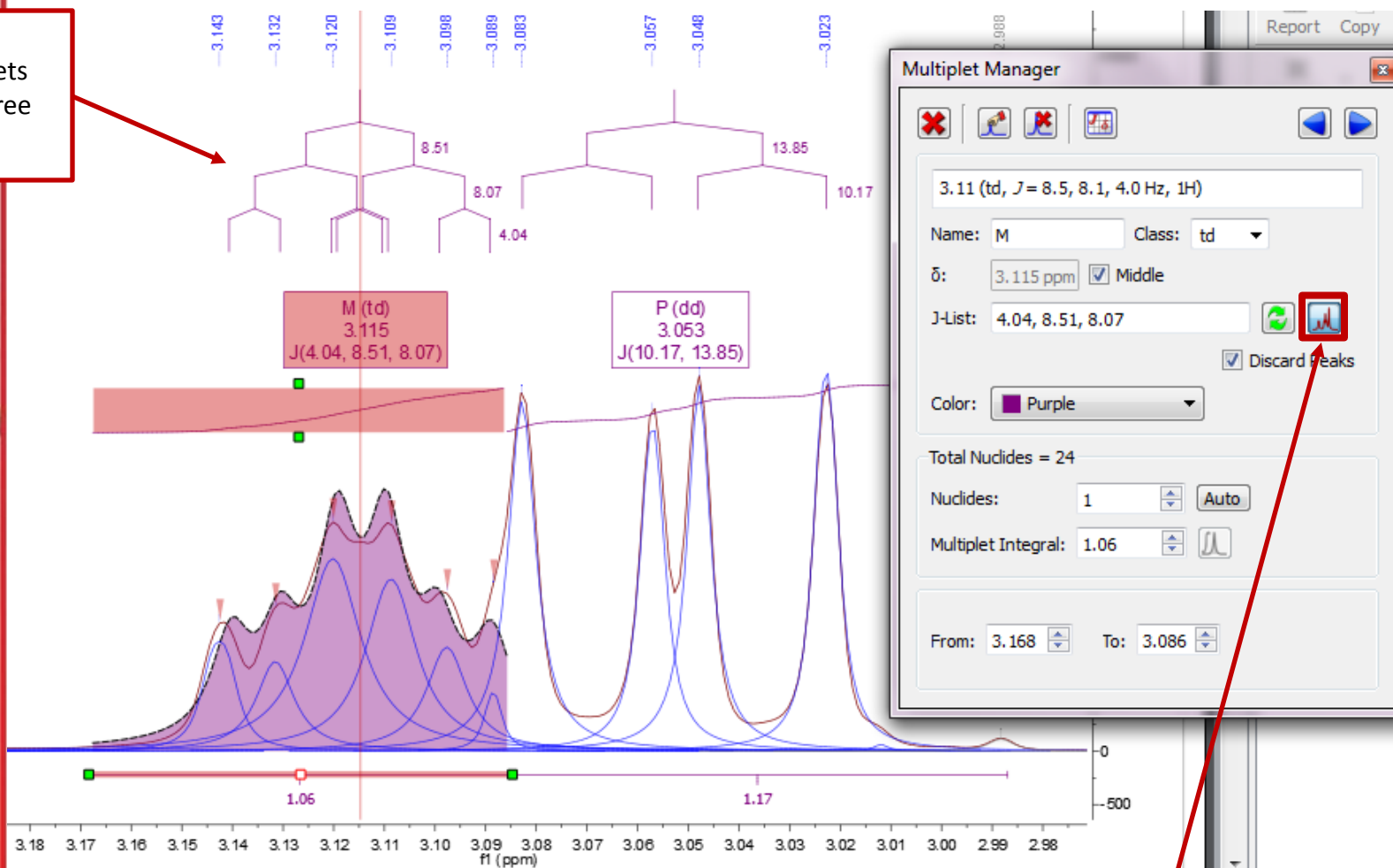
*Tip: You can also change the display of deconvolution peak curves in the Properties Dialog > Peaks > Curve tab.*

# To split partially overlapping multiplets (2)

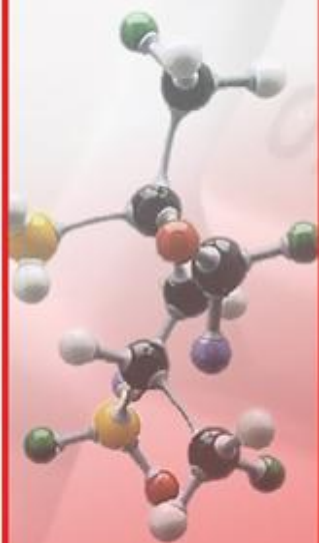


# Tools for verifying multiplet analysis results

Choose View > Properties > Multiplets and turn on the J's Tree option




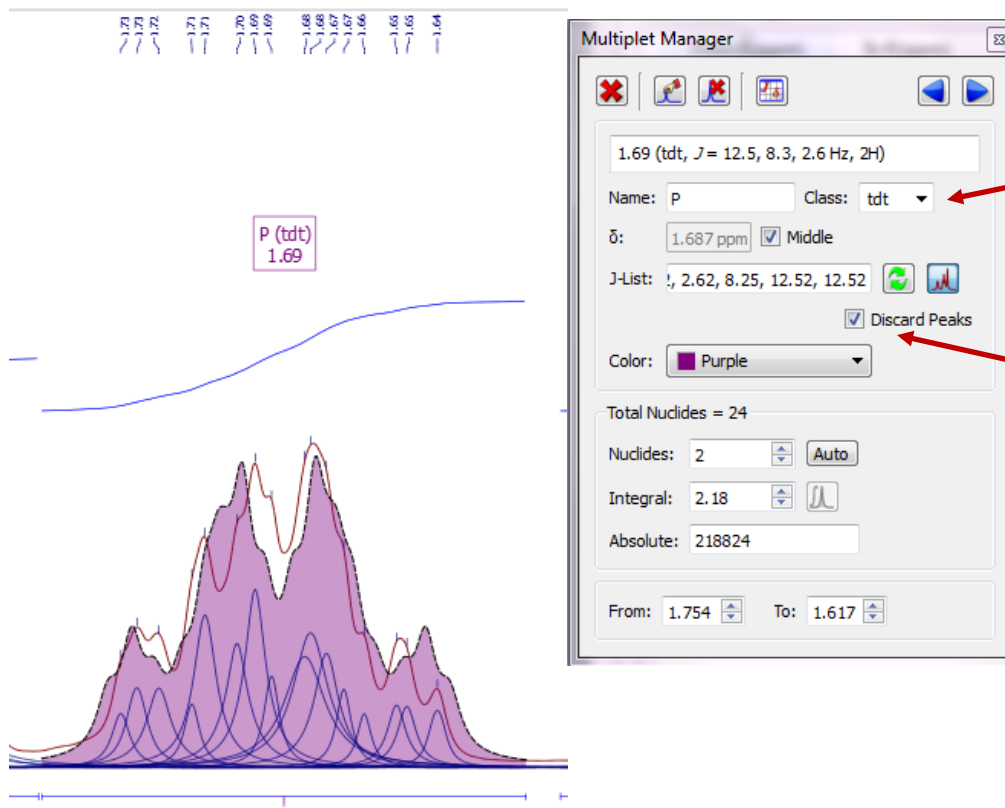
Use the simulation tool in the Multiplet Manager to simulate the multiplet and compare



# M

## To override the multiplet results in Multiplet Manager

- You can override the analysis results of a multiplet in Multiplet Manager.
- In this example, the multiplet was over-fit as a “tdt”. The simulated multiplet does not agree with the observed spectrum and hence it is wrong.
- Click  to turn off the simulated multiplet first. Select “m” from the drag-down menu of Class to override it.
- Or you can turn off the Discard Peaks option to include all peaks to the multiplet (and you get an “m” in this case).



Choose “m” from the drop-down menu to override the results

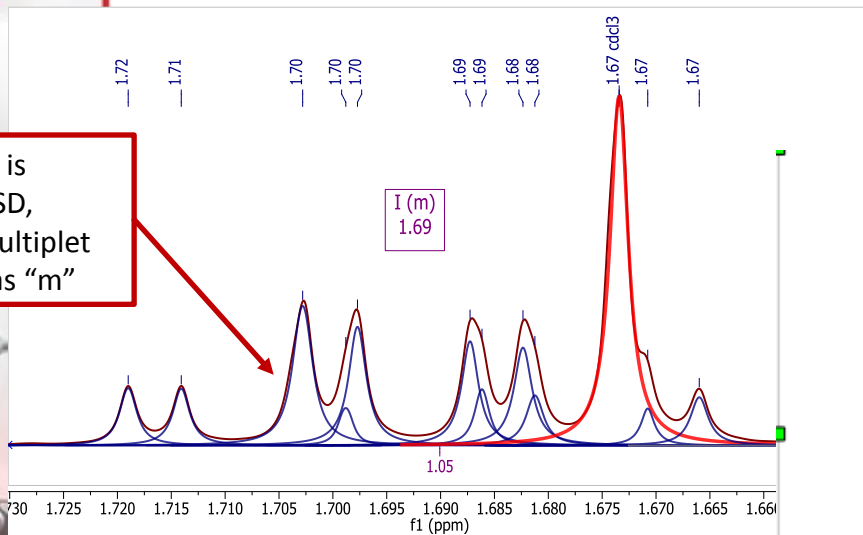
Or, you can turn off the Discard Peaks option to include all peaks and get a “m”



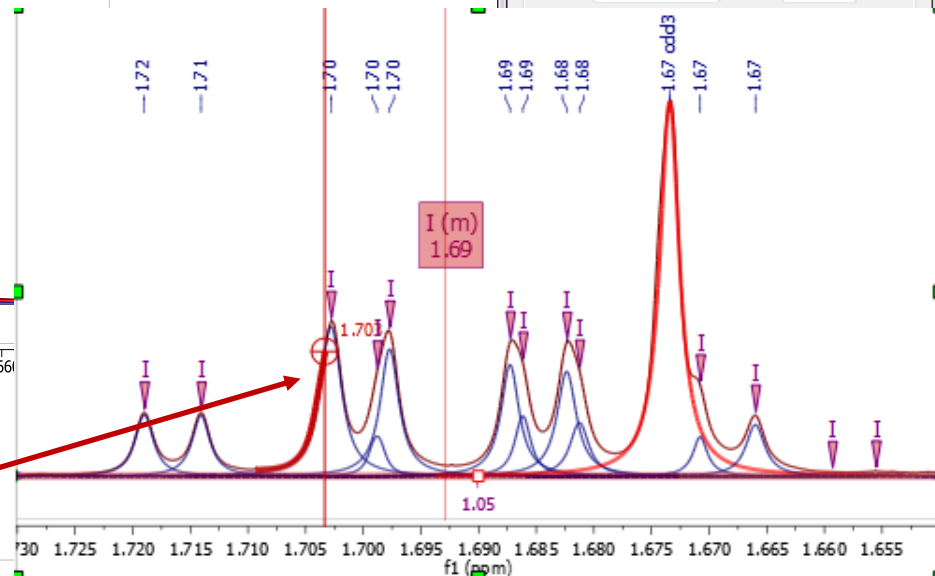
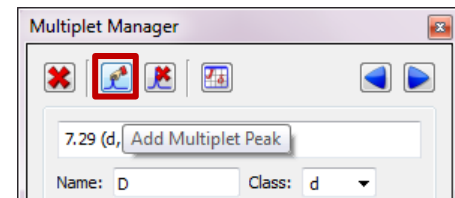
# To add a "missing" peak to a multiplet

# M

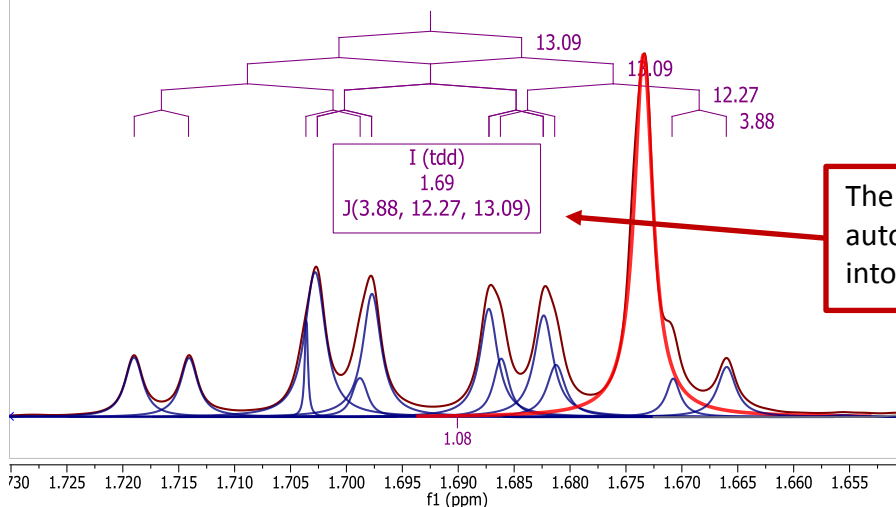
A small peak is missed by GSD, hence the multiplet is classified as "m"



Click the Add Multiplet Peak tool in the Multiplet Manager, click **SHIFT** key once, and click around here to add a shoulder peak



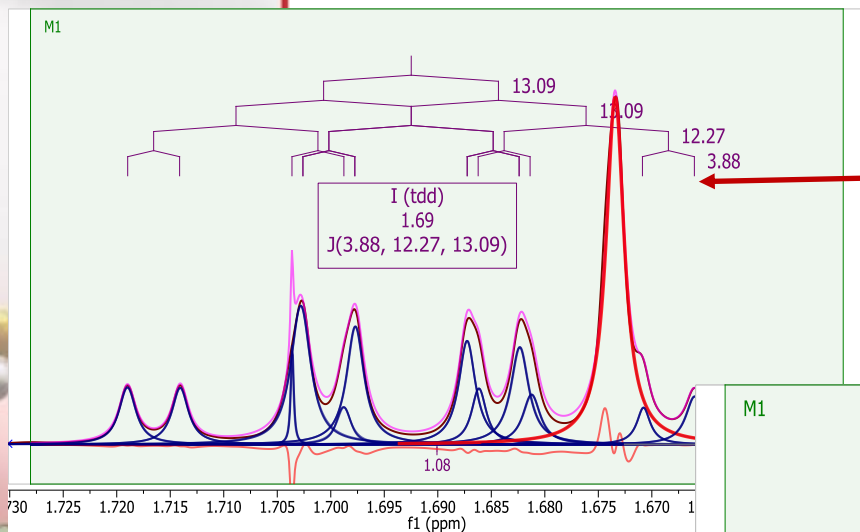
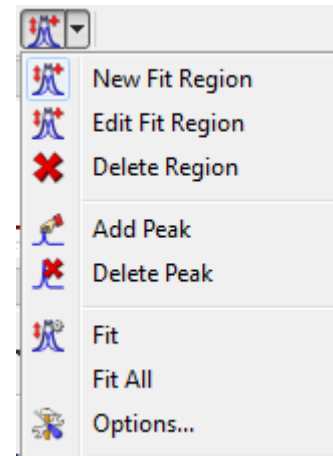
The added peak is automatically included into a "tdd" multiplet



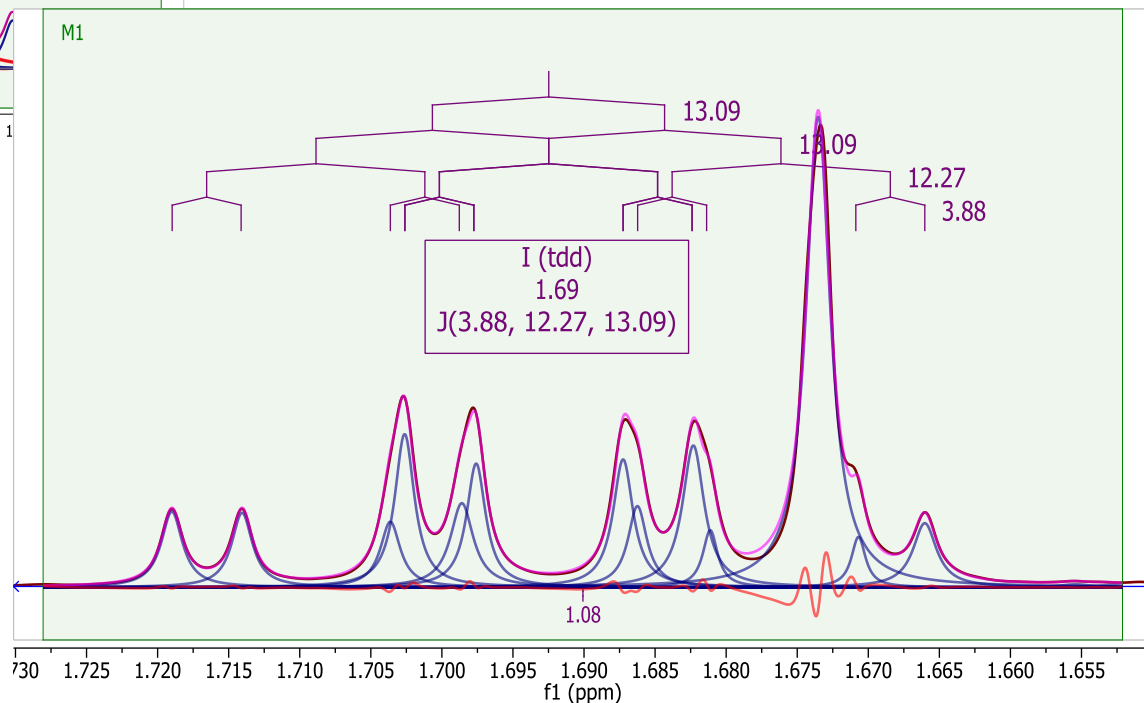


# Use Line Fitting to optimize peaks locally

- In addition to the novel GSD, Mnova has also the classic Line Fitting for fitting peaks locally, starting from initial peaks.
- The GSD and manually added peaks can be optimized as follows




Choose New Fit Region to define a region to fit.  
Choose Fit to fit the initial peaks to the spectral curve.  
Turn off the display of the GSD peak curves



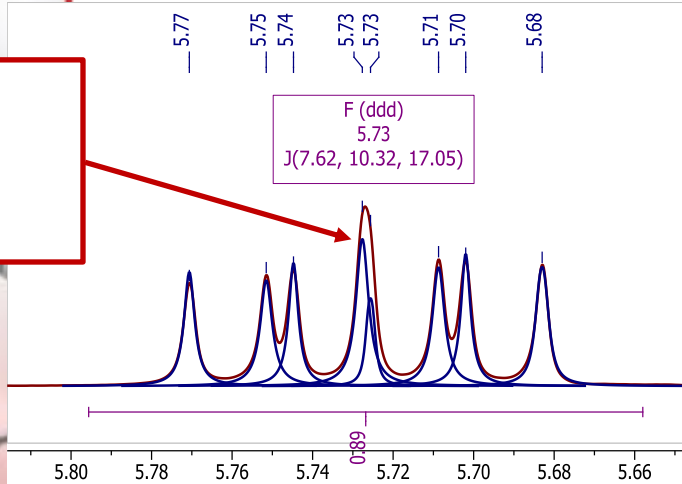
Note: The Line Fitting changes the GSD peaks and the relevant multiplet analysis results are automatically updated.



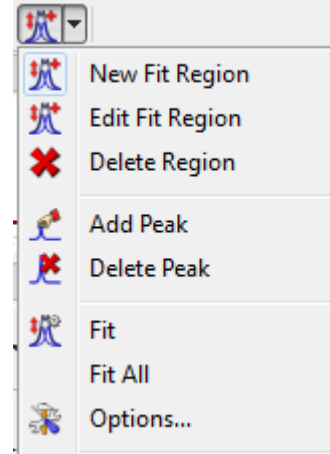
# Use Line Fitting to manually revise GSD peaks

 You can use the Line Fitting tools to manually edit the GSD peaks

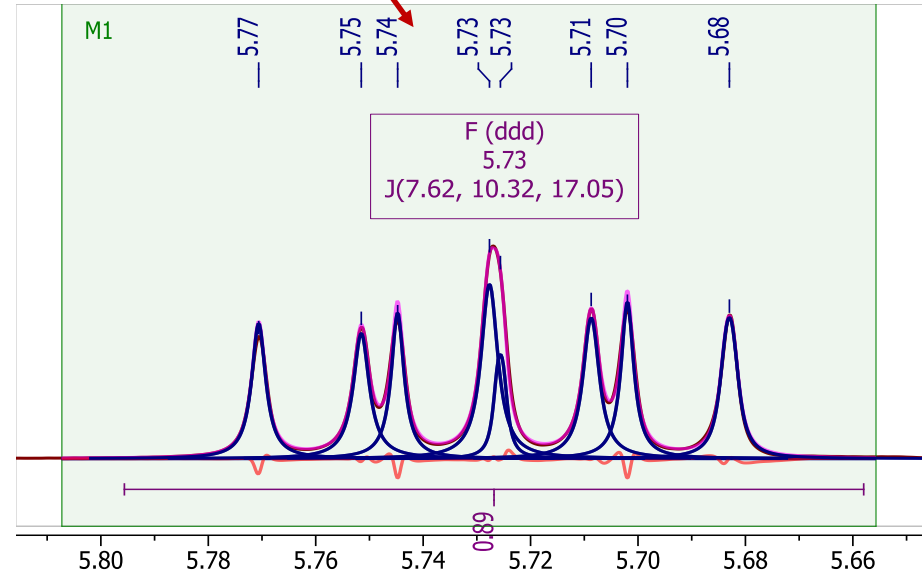
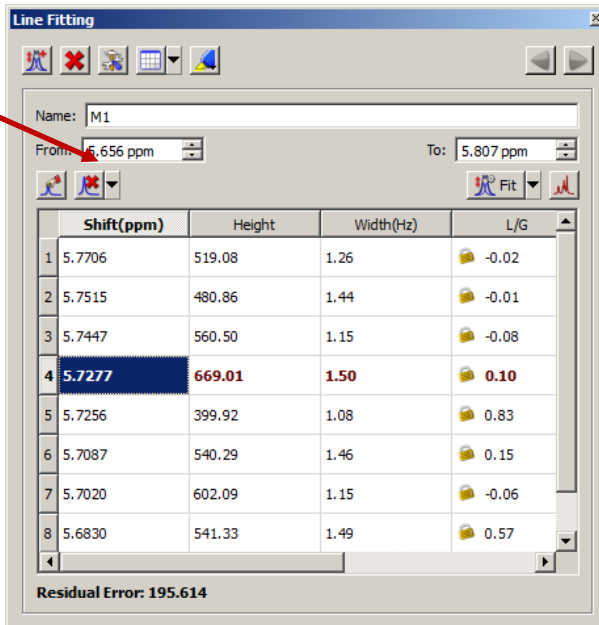
The two GSD peaks in the middle do not have the right heights.



Choose New Fit Region, click and drag to add a new fit region. The covered GSD peaks are taken as initial peaks; the simulated and residual curves are displayed



Choose **Edit Fit Region** to open the Line Fitting dialog



# Use Line Fitting to manually revise GSD peaks (2)

# M

Use the green handlers to select and edit a peak:

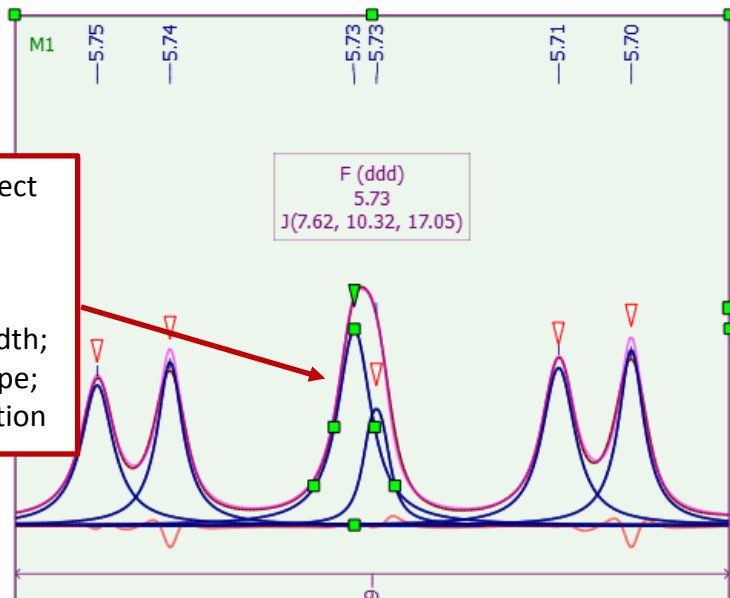
**Top triangle:** select the peak;

**Top box:** change the height;

**Middle boxes:** change the width;

**Lower boxes:** change the shape;

**Bottom box:** change the location



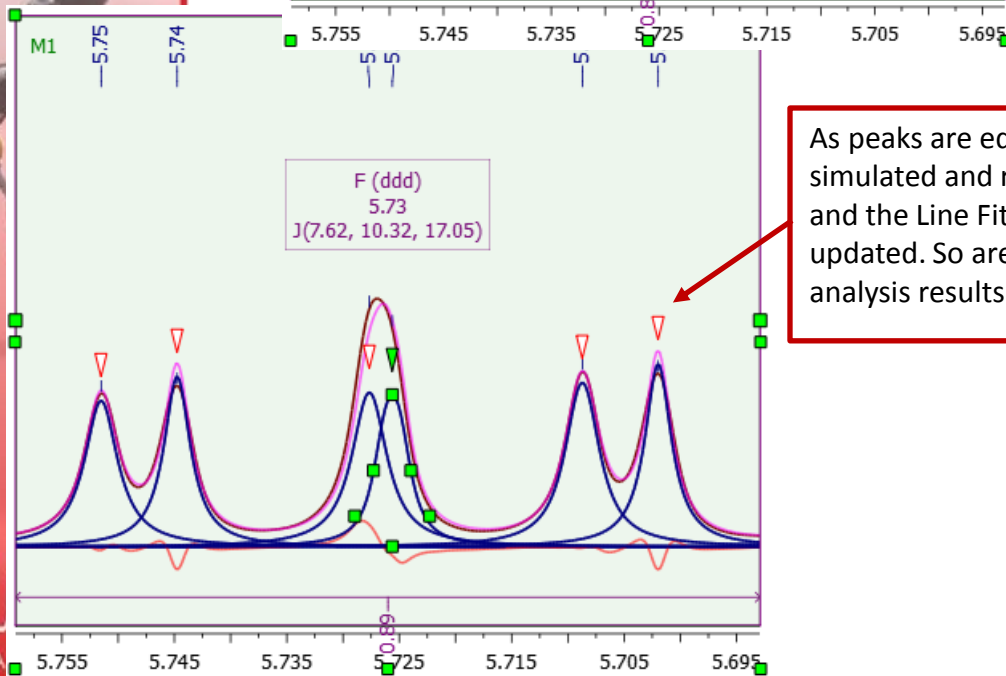
Line Fitting

Name: M1

From: 5.656 ppm To: 5.807 ppm

	Shift(ppm)	Height	Width(Hz)	L/G
1	5.7706	519.08	1.26	-0.02
2	5.7515	480.86	1.44	-0.01
3	5.7447	560.50	1.15	-0.08
4	5.7277	507.89	1.50	0.10
5	<b>5.7256</b>	<b>497.79</b>	<b>1.33</b>	<b>0.83</b>
6	5.7087	540.29	1.46	0.15
7	5.7020	602.09	1.15	-0.06
8	5.6830	541.33	1.49	0.57

Residual Error: 403.782



As peaks are edited, the simulated and residual curves and the Line Fitting table are updated. So are the multiplet analysis results

# Use Line Fitting to manually revise GSD peaks (3)

Line Fitting

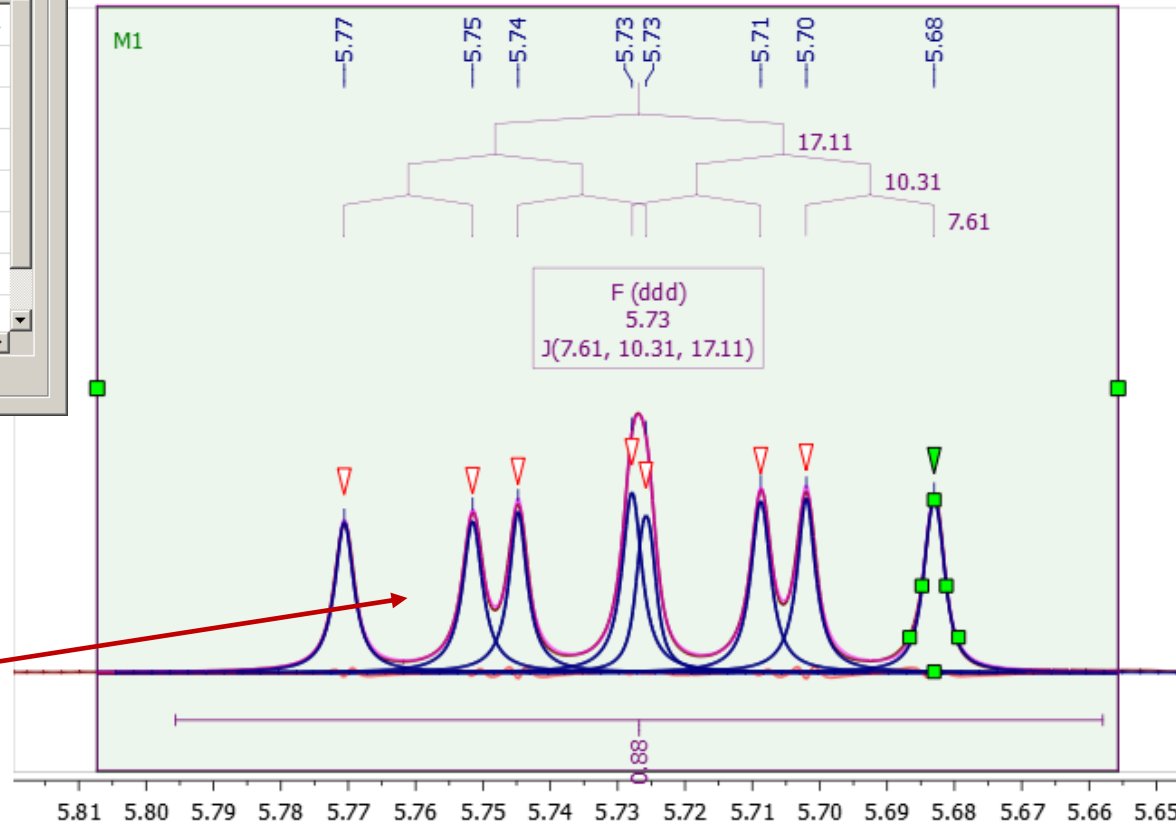
Name: M1

From: 5.650 ppm To: 5.816 ppm


	Shift(ppm)	Height	Width(Hz)	L/G	Area
1	5.7706	464.36	1.44	0.81	10079.254
2	5.7515	479.23	1.48	0.95	11186.631
3	5.7448	488.07	1.38	0.65	9653.496
4	5.7278	579.10	1.52	0.94	13933.007
5	5.7258	460.59	1.34	0.43	8119.618
6	5.7087	546.65	1.45	0.92	12420.826
7	5.7020	530.20	1.40	0.66	10619.343
8	5.6830	546.32	1.45	0.75	11703.136

Residual Error: 209.459

Click Fit to automatically optimize the current peaks and reach a better fitting to the experimental curve

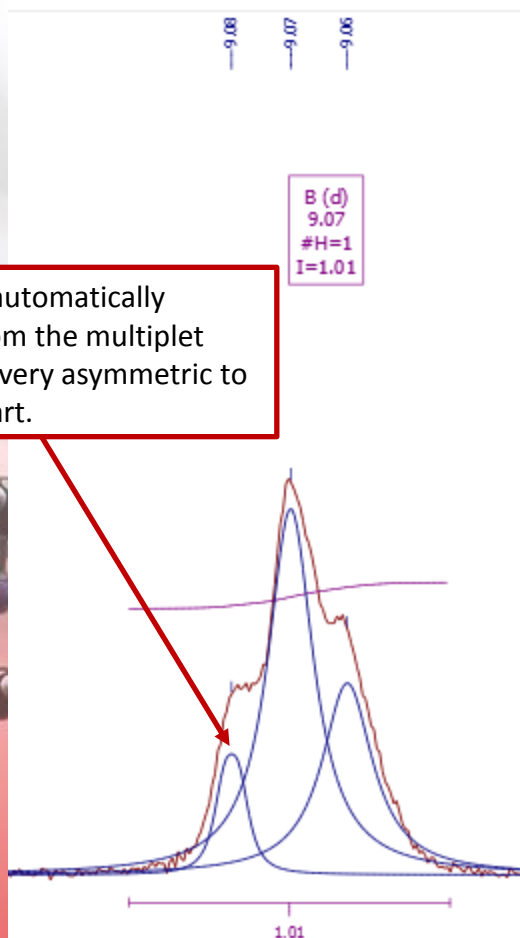


Note the change of the Residual Error, the simulated and residual curves. The multiplet analysis results are also automatically updated.

Tip: GSD uses a different line shape convention (called "Generalized Lorentzian" or "Kurtosis") than the normal Lorentzian-Gaussian (L/G) shapes used in Line Fitting. Click the Options Tool  to make sure that the Shape Types is set to "Lorentzian-Gaussian" if you want to convert GSD peaks to Lorentzian-Gaussian shapes during the fitting.

# To restore a discarded peak from a multiplet

# M



This peak is automatically discarded from the multiplet because it is very asymmetric to its counterpart.

Multiplet Manager

9.07 (t, 1H)

Name: B Class: t

$\delta$ : 9.071 ppm  Middle

J-List: 5.64, 5.64

Discard Peaks

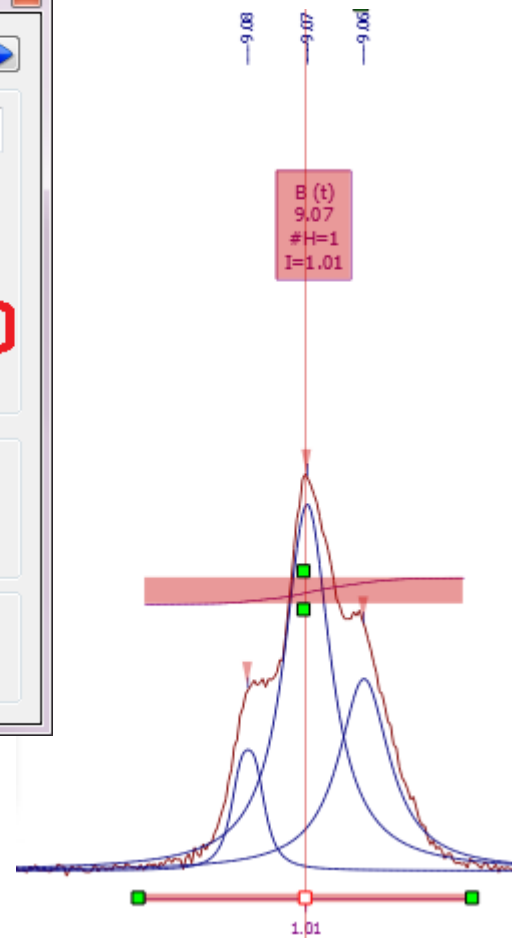
Color: Purple

Total Nuclides = 30

Nuclides: 1 Auto

Multiplet Integral: 1.01


From: 9.097 To: 9.045



Turn off the Discard Peaks. All the 3 peaks are taken as parts of the multiplet, and they are classified as a triplet.



# To re-assign peaks to multiplets

- If a peak is assigned to a wrong group, use the Add Multiplet Peak tool  in the Multiplet Manager to re-assign it to a different group
- In the following example two peaks were re-assigned, forming a different pair of doublets:

Multiplet Manager

7.29 (d) Add Multiplet Peak

Name: D Class: d

$\delta$ : 7.290 ppm  Middle

J-List: 2.70

Discard Peaks

Color: Purple

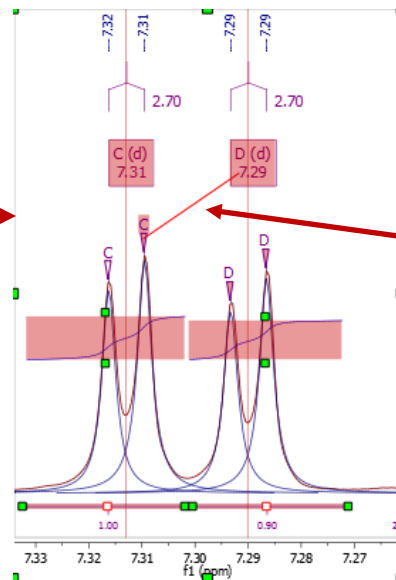
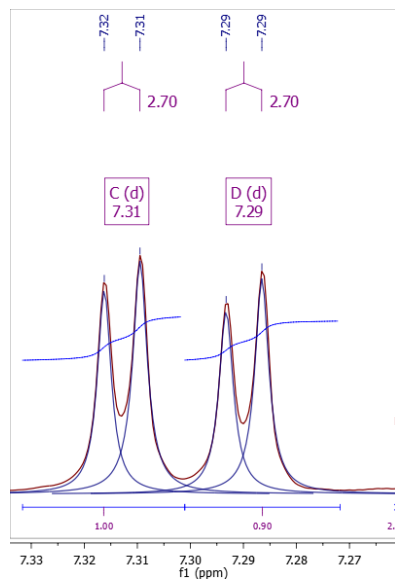
Total Nucleides = 4

Nucleides: 1

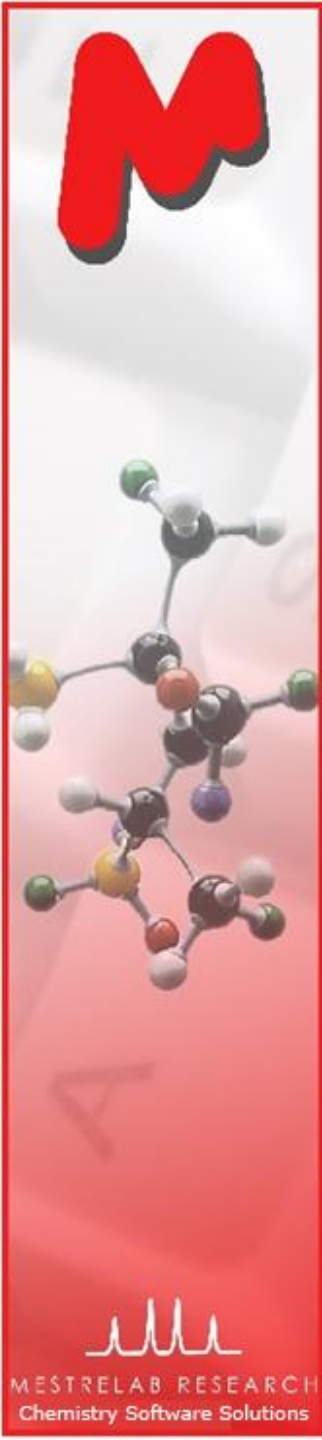
Integral: 0.90

Absolute: 40196.1

From: 7.301 To: 7.272



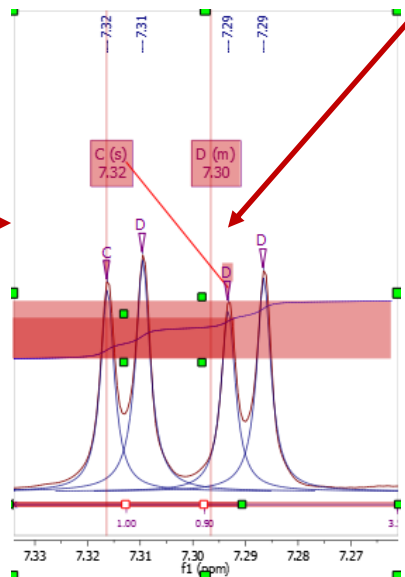
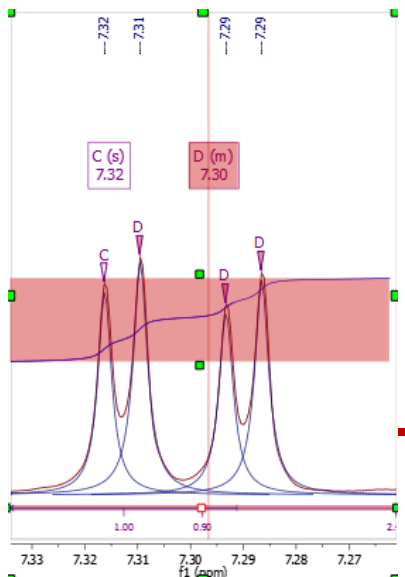
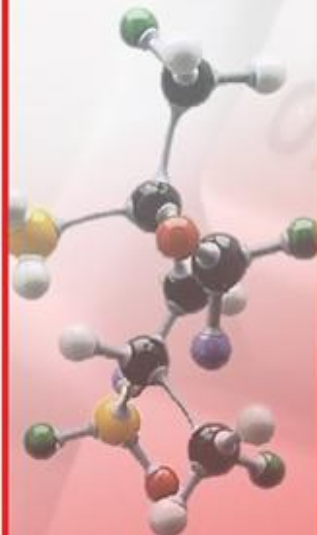
Click on the triangle mark on top of the peak, drag it to the multiplet label "D" to assign it to a different group



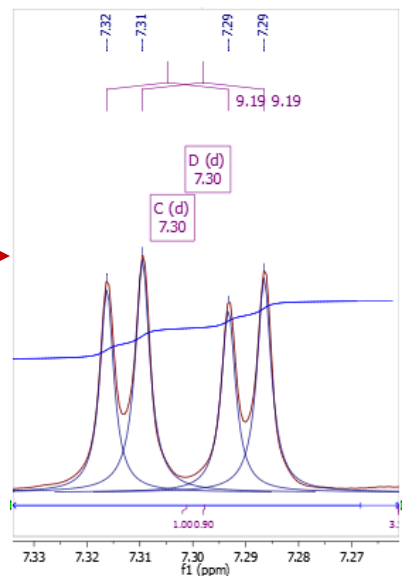


# To re-assign peaks to multiplets (2)

# M



Click on the triangle mark on top of the peak, drag it to the multiplet label "C" to assign it to Mutliplet "C"

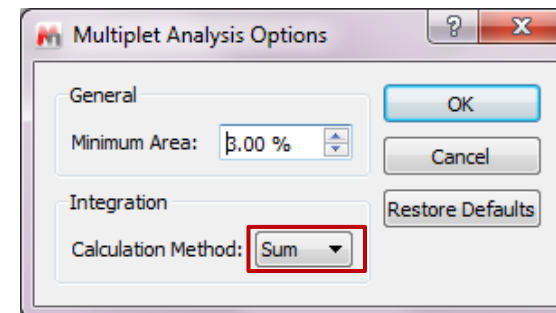
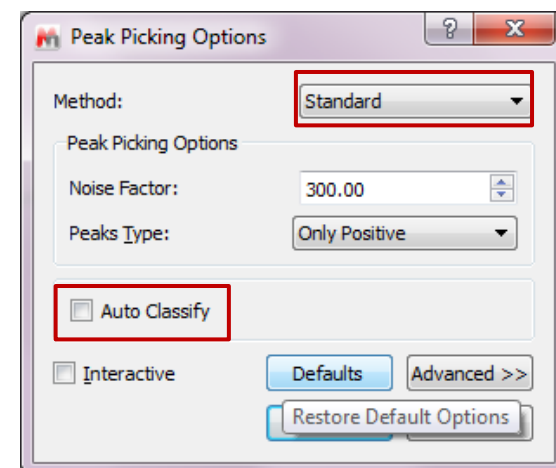





# M

## Change the settings to traditional multiplet analysis

- If you do not like the GSD-based peak picking and multiplets analysis, you can change the options back to the traditional ways.
- Open a 1D NMR, then do the following to turn off the use of GSD-based peak picking and multiplet integration:
  - For **Peak Picking Options**, change the **Method** to **Standard** to use the traditional peak maxima-based method, also turn off the **Auto Classify** if you don't want to classify peak types automatically
  - For **Multiplet Analysis Options**, change the **Calculation Method** to **Sum**\*

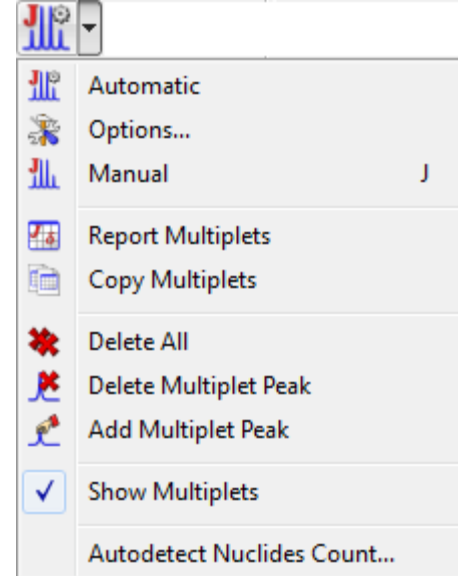


\* Note: The results from Integration  is independent of those from the Multiplet Integration. So only the Peak Picking options and Multiplet Analysis options will affect the multiplet analysis results. Sum is the traditional method of integration by summing up all points within the integration region.



# To report multiplets

- Click **Report Multiplets** to report the results in a journal format:
- To change journal format: choose **View | Tables | Multiplets** to display the Multiplets Table. Click **Setup Report**



**Multiplets**

Report Multiplets Copy Multiplets **Setup Report** Delete

<sup>1</sup>H NMR (400 MHz, CDCl<sub>3</sub>) δ 8.62 (d, J = 4.5 Hz, 1H), 7.95 (d, J = 9.2 Hz, 1H), 7.51 – 7.43 (m, 1H), 7.30 (dd, J = 9.2, 2.7 Hz, 1H), 7.21 (d, J = 2.7 Hz, 1H), 5.73 (ddd, J = 17.1, 10.3, 7.6 Hz, 1H), 5.48 (d, J = 4.4 Hz, 1H), 4.99 – 4.85 (m, 2H), 3.87 (s, 3H), 3.44 – 3.31 (m, 2H), 3.17 – 2.99 (m, 2H), 2.69 – 2.56 (m, 2H), 2.30 – 2.18 (m, 1H), 1.90 (s, 2H), 1.83 – 1.62 (m, 3H), 1.61 – 1.34 (m, 2H).

	Name	Shift	Range	H's	Integr
1	C (m)	7.46	7.51 .. 7.43	1	0.99
2	A (d)	8.62	8.67 .. 8.58	1	0.89
3	C (m)	1.71	1.83 .. 1.62	3	3.26

**Multiplet Report**

JACS

All as Ranges

m's as Ranges

Ascending Order of Shifts

Report Js

Reduce J List

Use Extended Solvent Names

OK Cancel


**Multiplet Report**

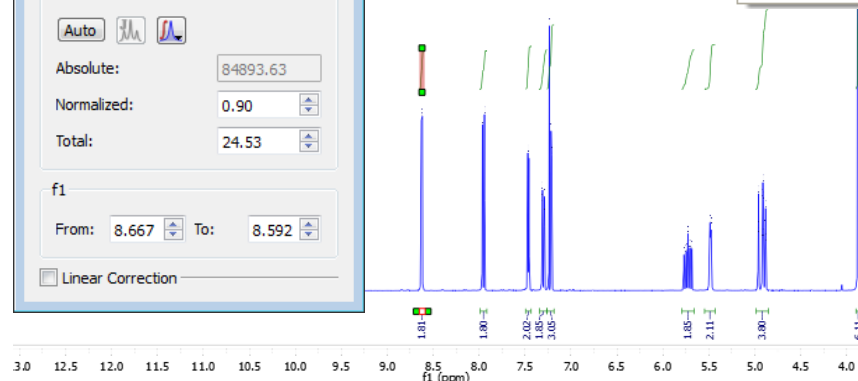
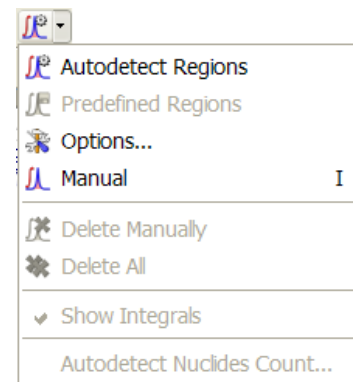
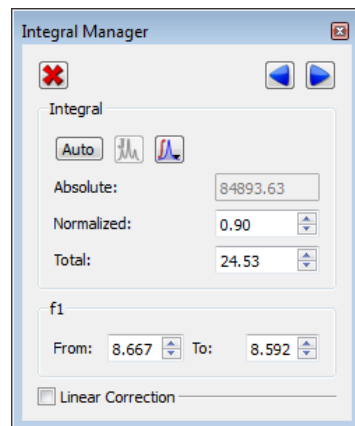
JACS

- Angewandte
- JACS
- J.Med.Chem
- J.Nat.Products
- Japanese Patent
- Organometallics
- Polyhedron
- RSC
- Tetrahedron
- Tetrahedron Letters
- US Patent

*Tip: From the Multiplet Table, click **Copy Multiplets** and then paste the texts to your document. Click **Copy Table** and then paste the spreadsheet to your document. The table can be customized using **Setup Table**.*

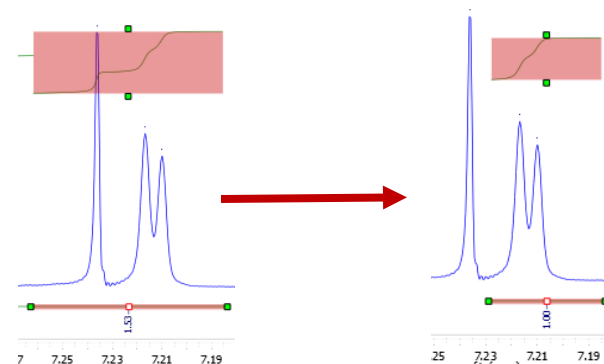
# To integrate peaks independent of multiplet analysis

- Click  to do auto integration or click **I** to do it manually
- Double click on an integral curve to popup Integral Manager:

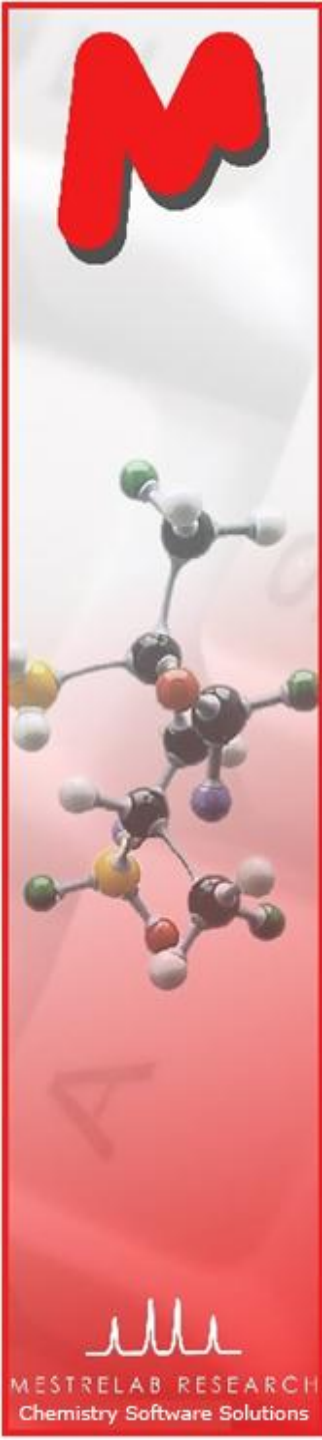


- Type a Normalized value to normalize the integrals
- Browse, delete, change, split integrals interactively if needed

Click and drag the left green box to change the range of the integral

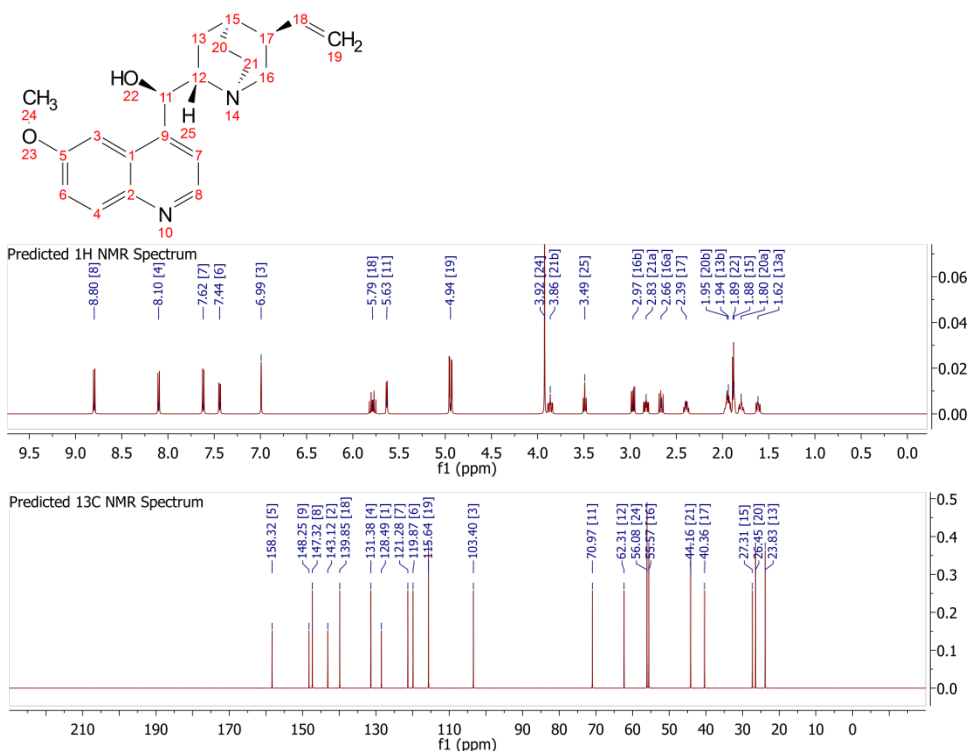
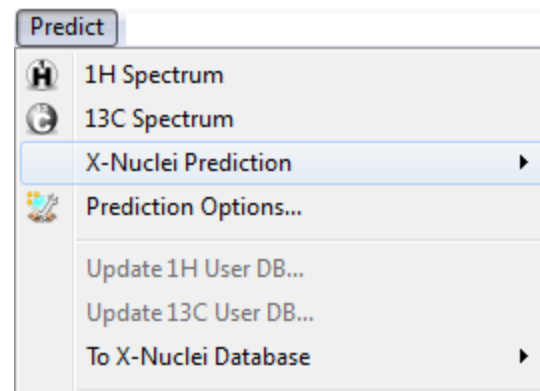


\* Note: The results from Integration is independent of those from the Multiplet Integration. Use Integration Options to change the method and other parameters.



# To predict NMR from a structure\*

- Open a new document (**File | New**) or a new page (**Edit | Create New Page**)
- Copy a structure from ChemDraw, Isis/Draw or ChemSketch, and paste to Mnova, or open a .mol, .cdx or a .sdf file
- Choose an command from the **Predict** menu

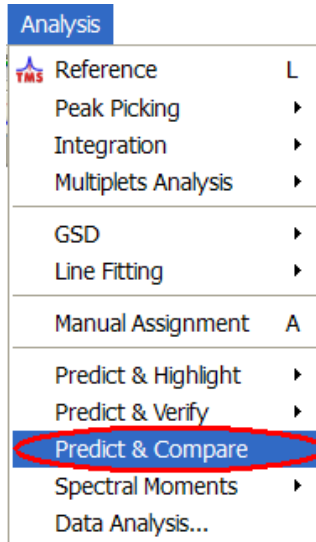


- Tips:*
- Choose **Molecules | Prediction Options** to change settings
  - You can turn on/off the atom numbers by right-clicking on the structure and choose **Properties**.
  - You can open the **Prediction Table** to list the predicted shifts and *J*-couplings, and manually change them.

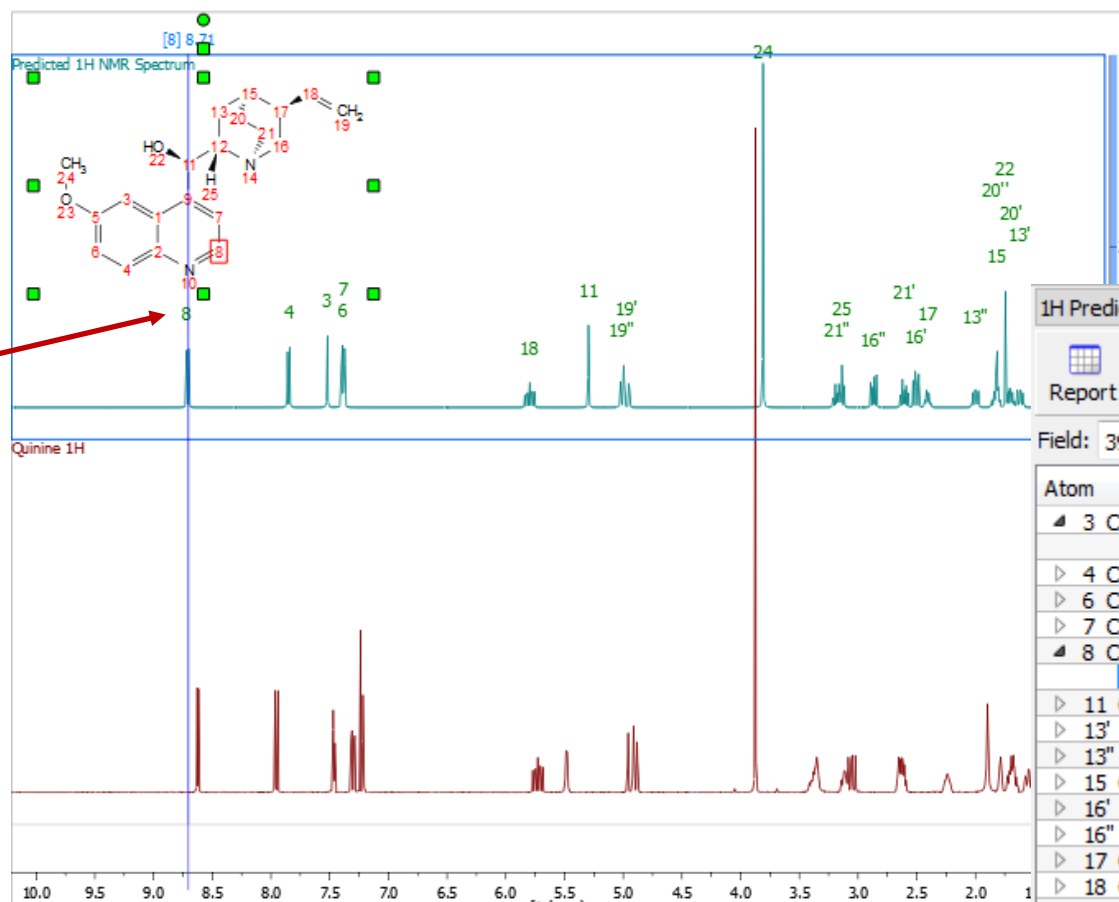
\* A separate license of Mnova NMRPredict Desktop is needed.

# To predict NMR & verify your structure

- Open your  $^1\text{H}$  (or  $^{13}\text{C}$ ) **spectrum** in a new page
- Copy your **structure** from ChemDraw or Isis/Draw
- Choose **Analysis | Predict & Compare**. The predicted spectrum is stacked with the experimental one for visual comparison



You can drag the label of a predicted peak to change its chemical shift. You can also change the predicted J-couplings in the 1H Prediction Table.



# To assign a 1D $^1\text{H}$ spectrum

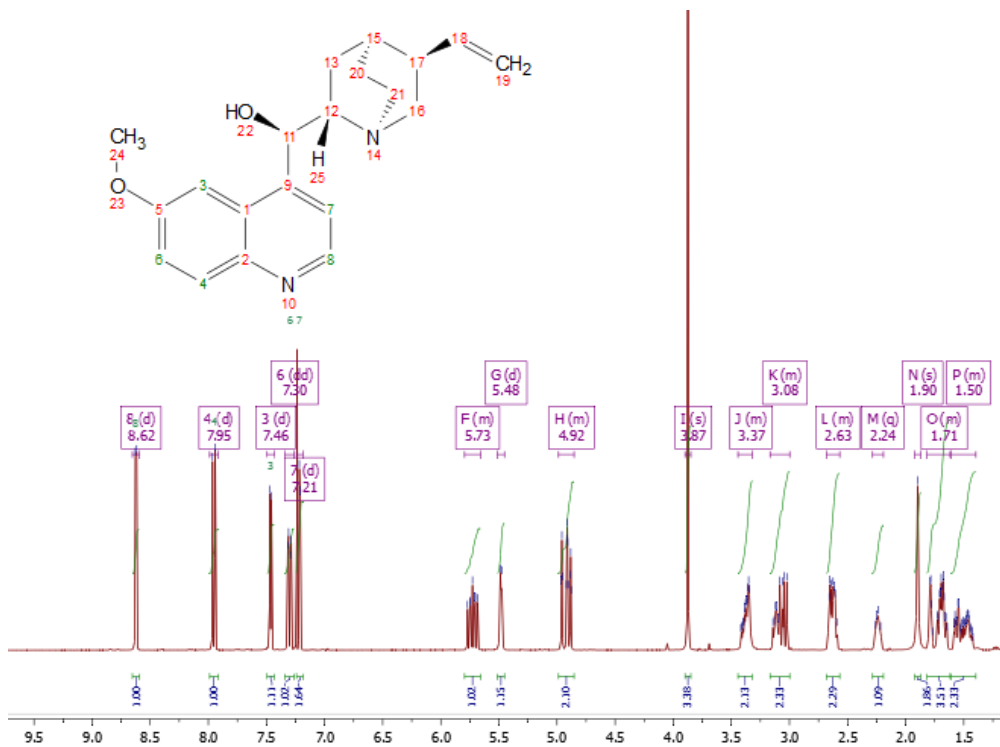
Click **A** key (or choose **Analysis | Manual Assignment**) to enter Assignment mode.



Click on an **atom** in the structure. Then choose the **peak** you want to assign. There are 3 ways to do it:

- A picked **multiplet**, by clicking on the multiplet label, or
- A **peak top**, or any point in the spectrum by clicking on it, or
- A **range** in the spectrum, by click-and-dragging to cover it

You can predict the  $^1\text{H}$  spectrum to assist your assignment\*

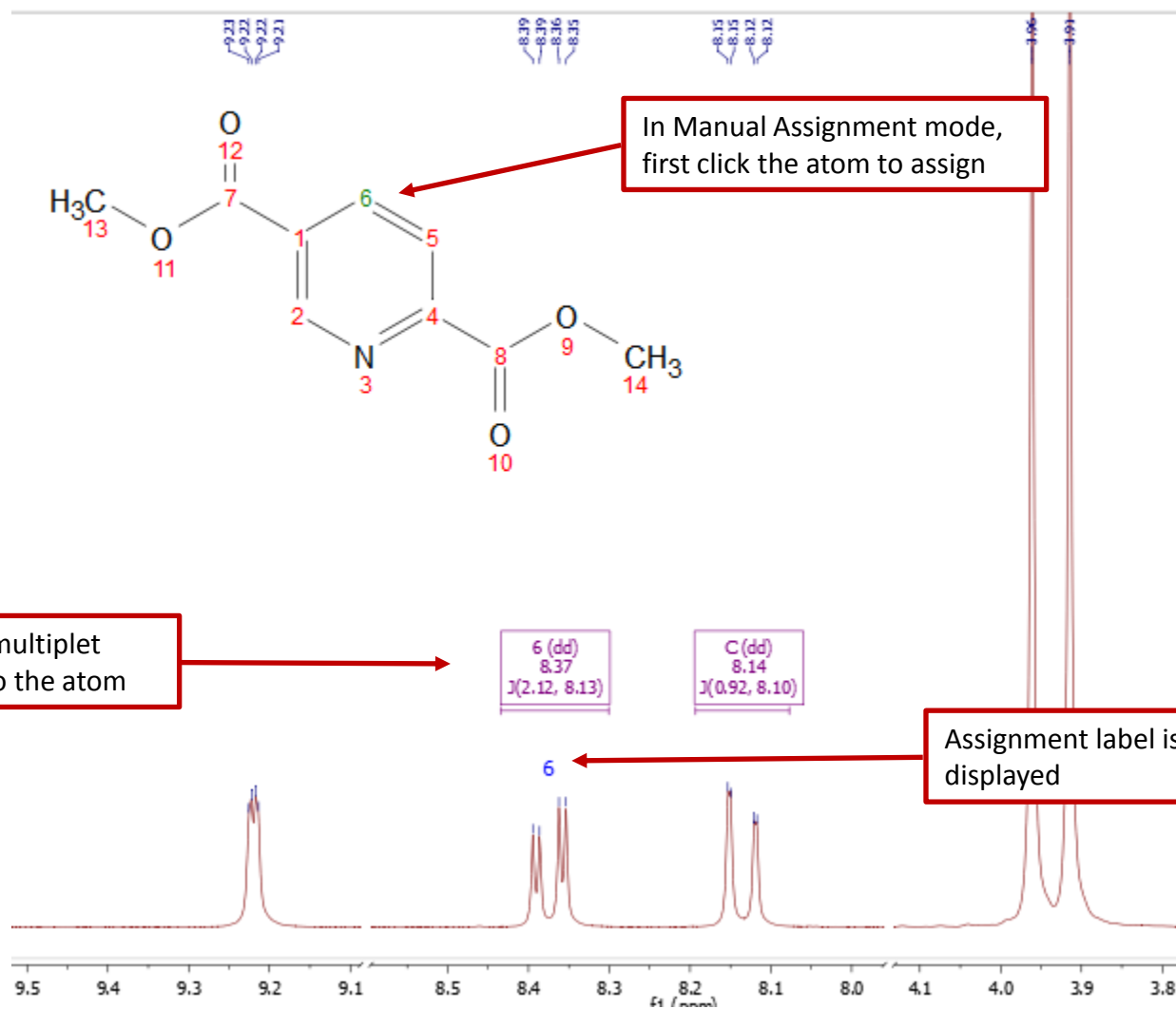


*\*Needs a separate license for Mnova NMRPredict Desktop*



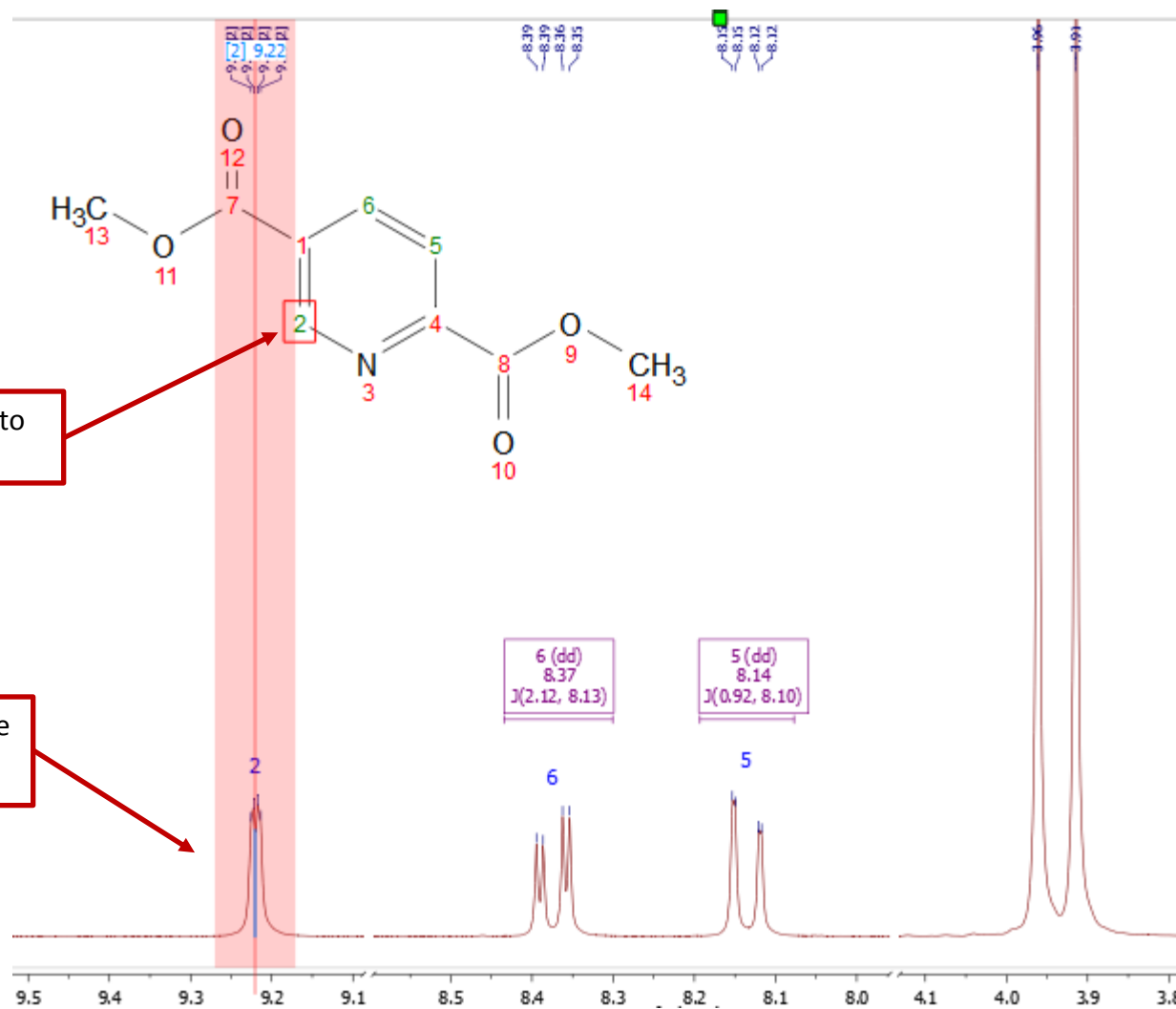


# To assign a multiplet to an atom



*Tip: After the assignment, the atom label is changed to green. The multiplet label shows the atom label. The multiplet label can be turned off by unchecking Analysis | Multiplet Analysis | Show Multiplets*

# To assign a region to an atom

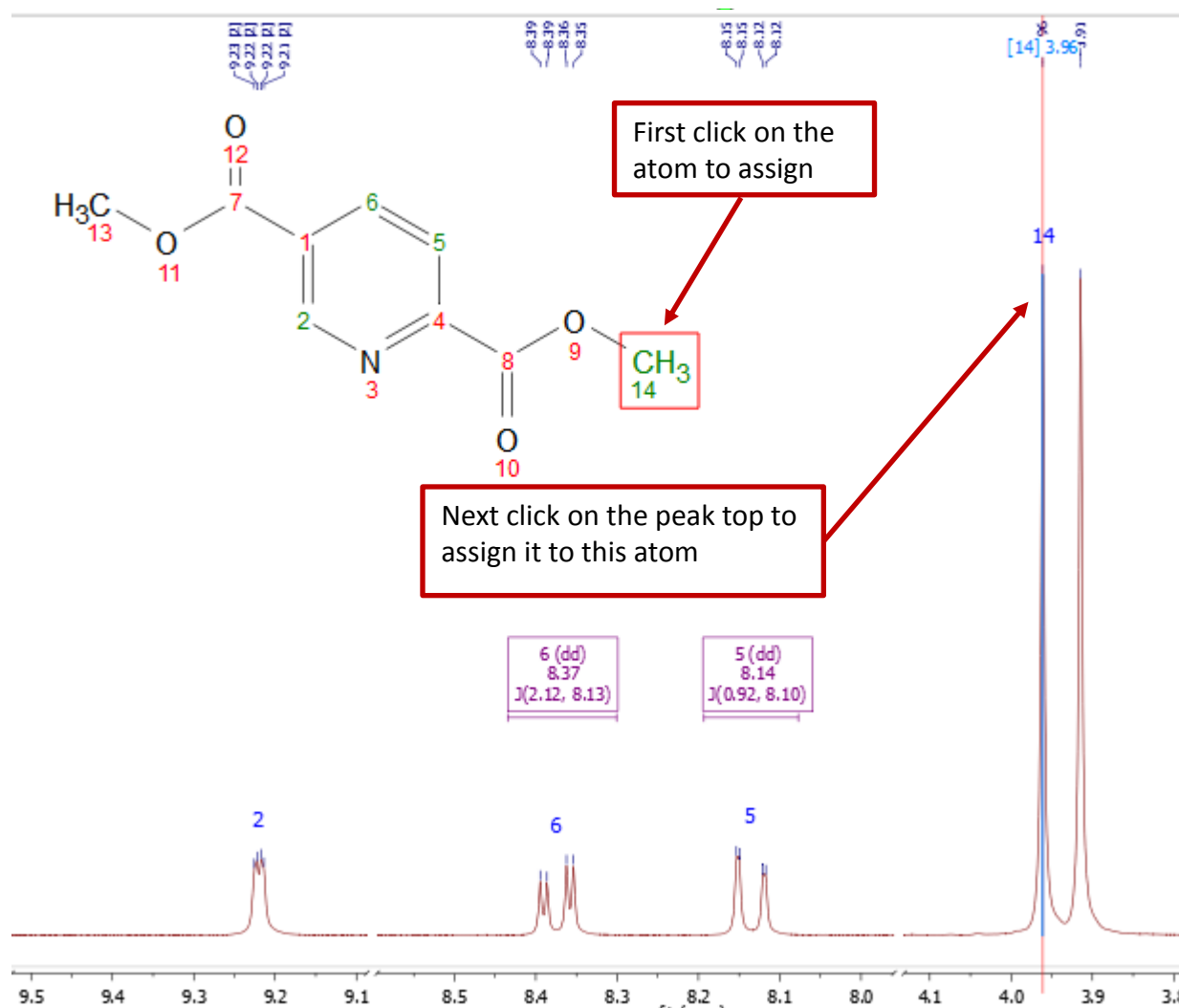


First click on the atom to assign

Next click-and-drag around the peak to assign it to the atom



# To assign a peak top to an atom

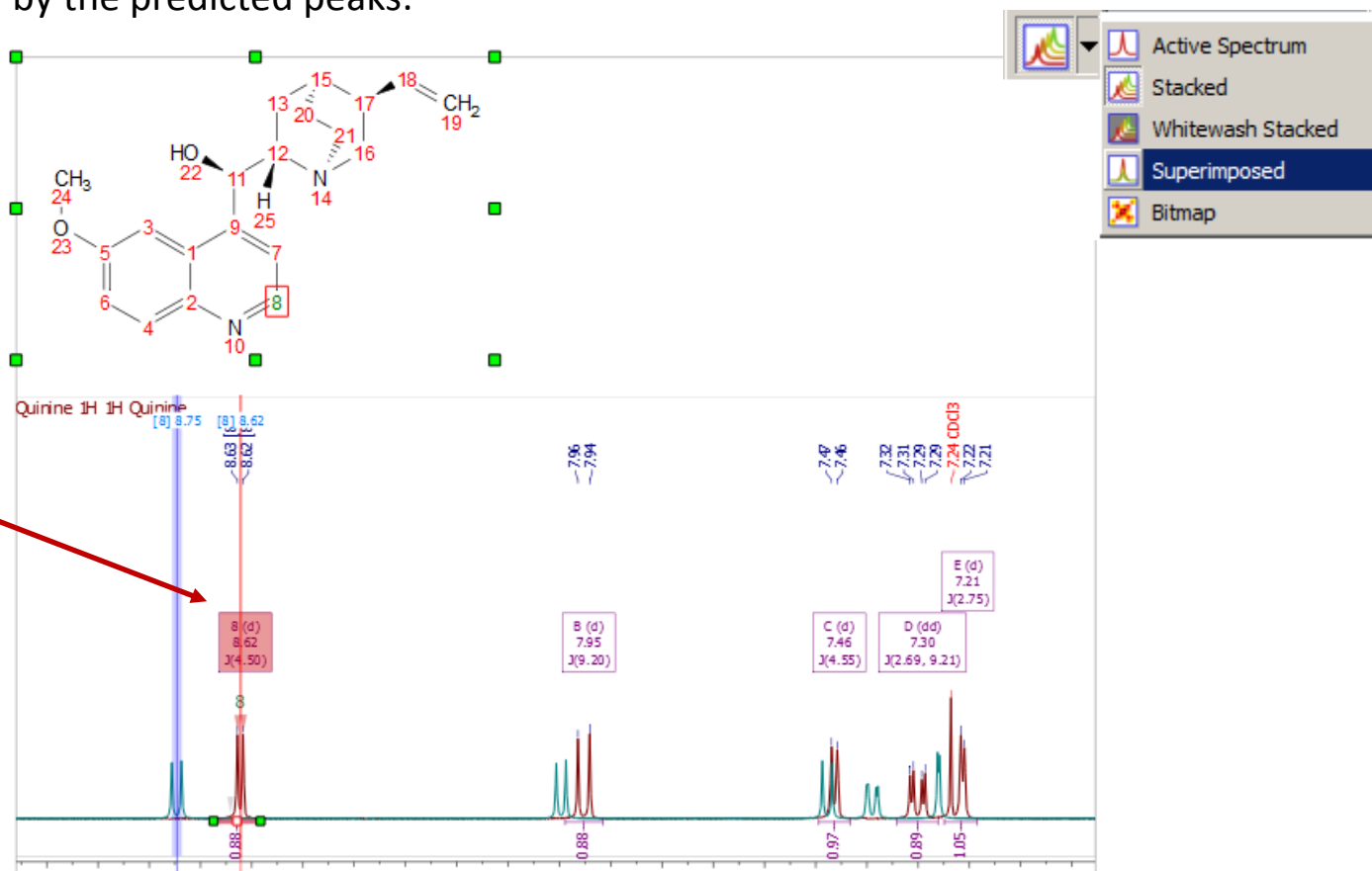


*Tip: By Default, Mnova automatically snaps to a peak top (with interpolation). Click **Shift** key one time to toggle it off if you want to choose a shoulder peak.*



# To predict NMR & help you assign peaks

- Open your  $^1\text{H}$  (or  $^{13}\text{C}$ ) **spectrum** in a new page, do multiplet analysis or peak picking as usual
- Copy your **structure** from ChemDraw or Isis/Draw
- Choose **Analysis | Predict & Compare**. The predicted spectrum is stacked with the experimental one for visual comparison
- Switch to **Superimposed Mode** so you can assign the multiplets/peaks guided by the predicted peaks:



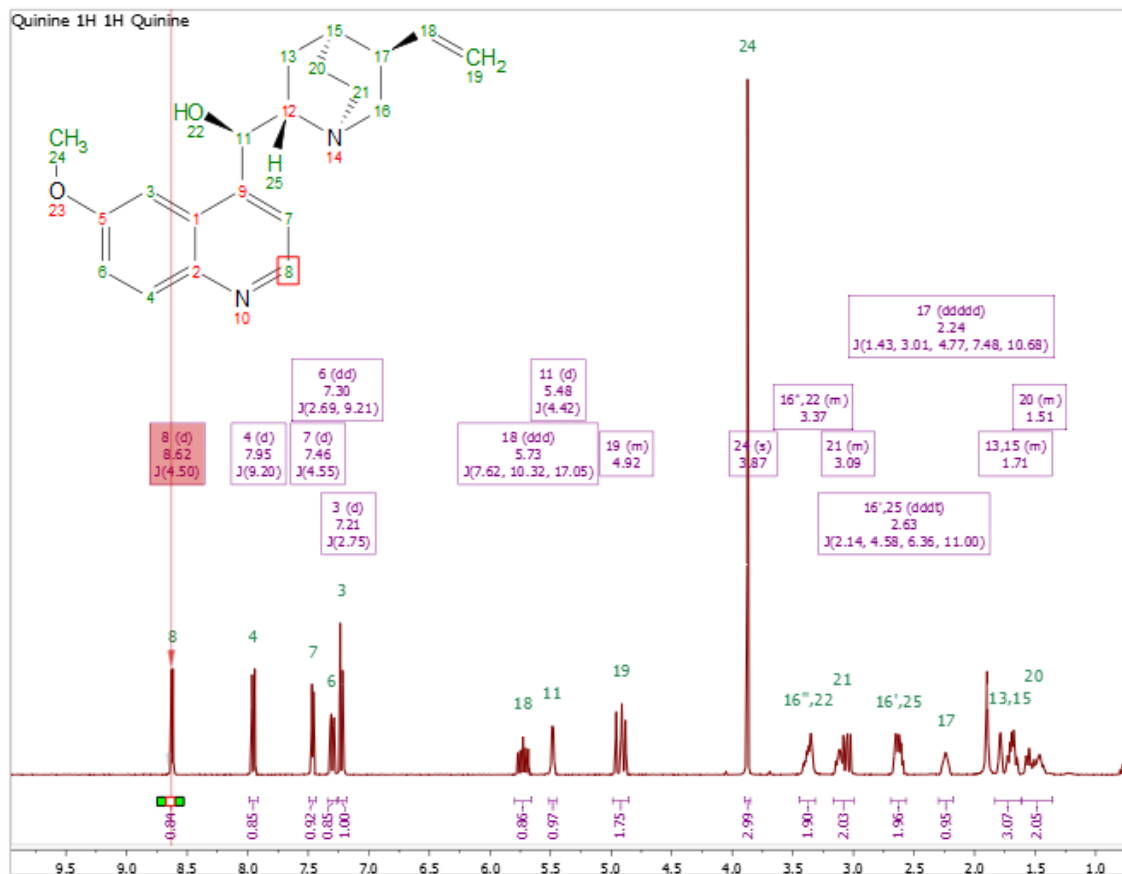
Use Shift + Up Arrow key to change the active spectrum and see the multiplet labels and predicted peak labels. In Assignment mode, click on a multiplet label and then an atom to make the assignment.

Blue: predicted peaks  
Red: observed peaks



# Automatic assignment of $^1\text{H}$ spectrum\*

- Open your  $^1\text{H}$  spectrum in a new page, copy your structure from ChemDraw or Isis/Draw
- Choose **Analysis | Automatic Assignment**. Mnova does multiplet analysis (if not done yet), predicts H-1 spectrum, and automatically assigns H-1 peaks

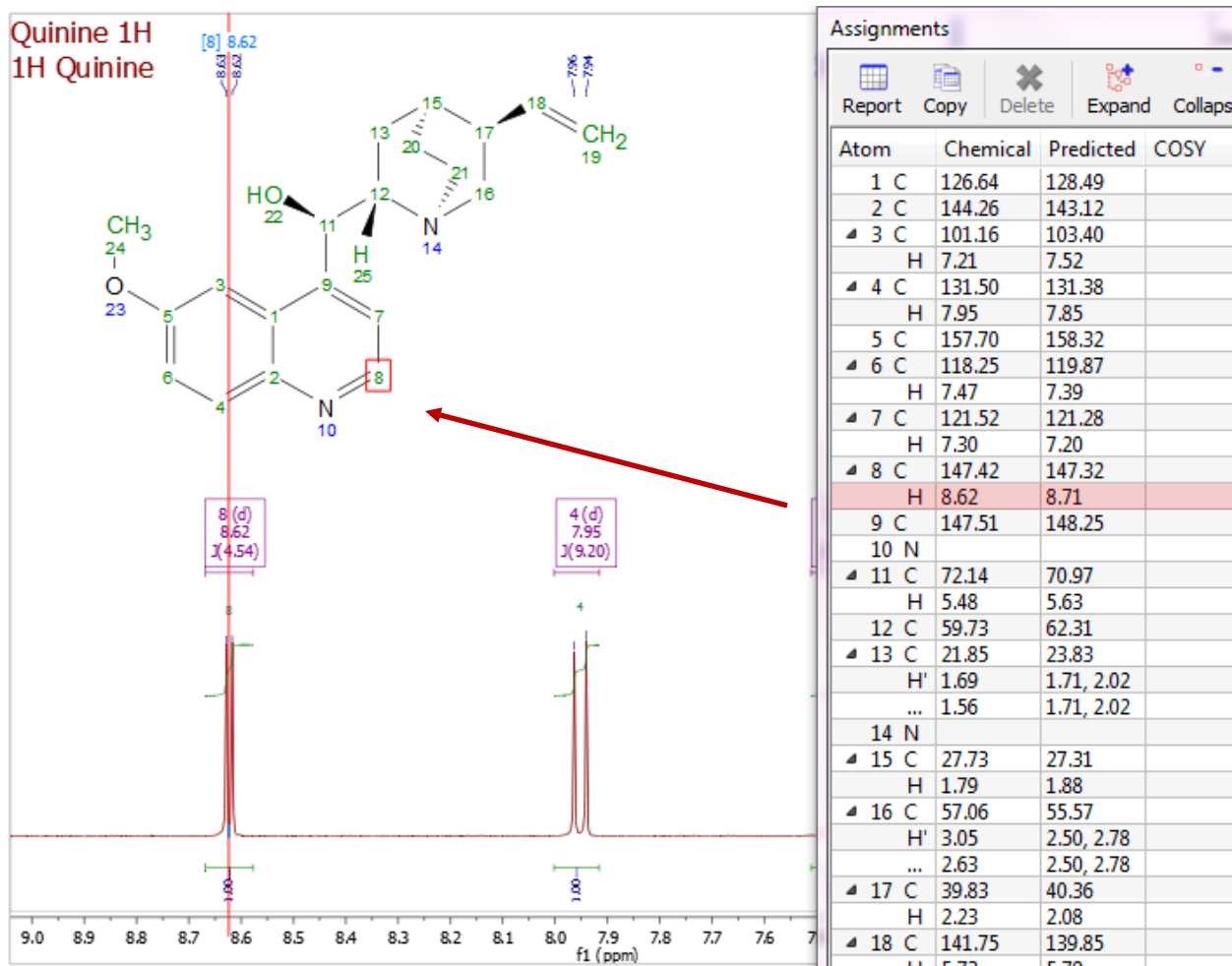


\*A separate license of Mnova NMRPredict Desktop is needed in addition to Mnova NMR.

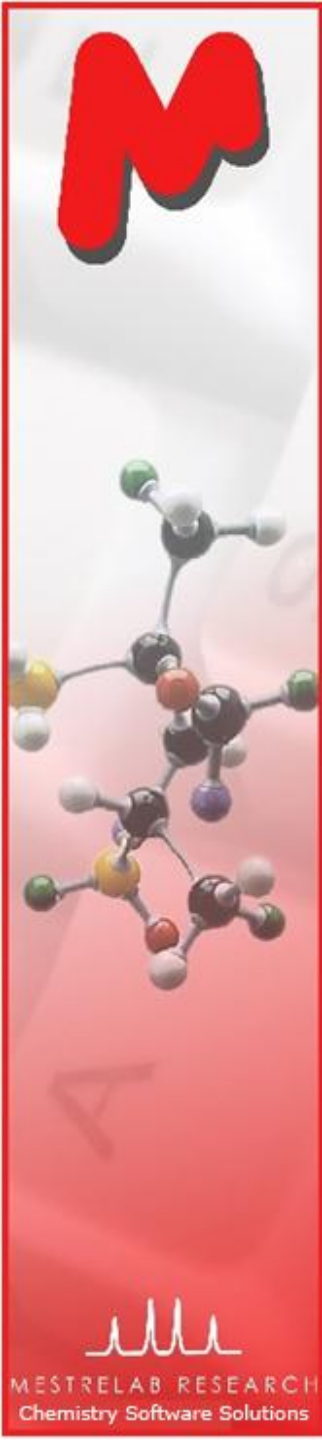
Tip: you can do multiplet analysis and clean them up prior to auto assignment. Also, try **Mnova Verify** that automatically verifies your proposed structures (<http://mestrelab.com/software/mnova-verify/>).

# To display and browse assignment results

- Choose **View | Tables | Assignments** to open the Assignments Table
- The Table and the structure are correlated: You can click a row to highlight the atom (and its assigned peak), and vice versa



\* You can right click on an atom and choose **Edit Atom Data** to change its label. Changed labels will be used in Assignments Table and other relevant reports.



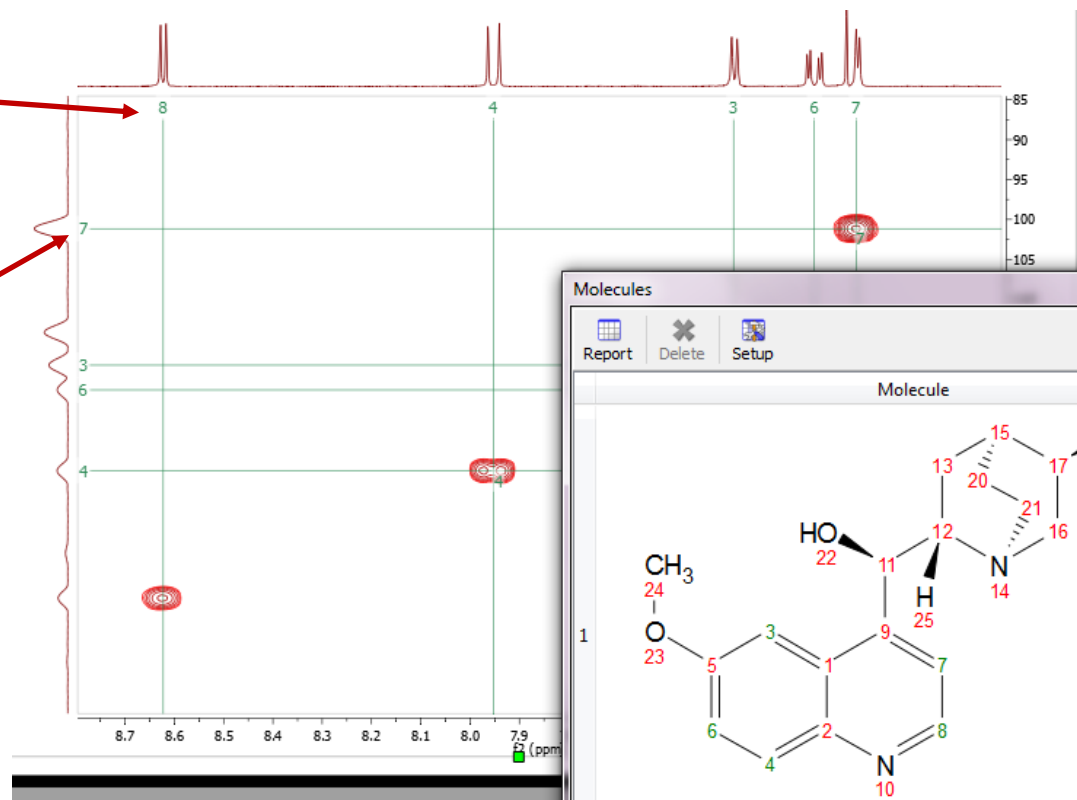


# If you have 2D HSQC

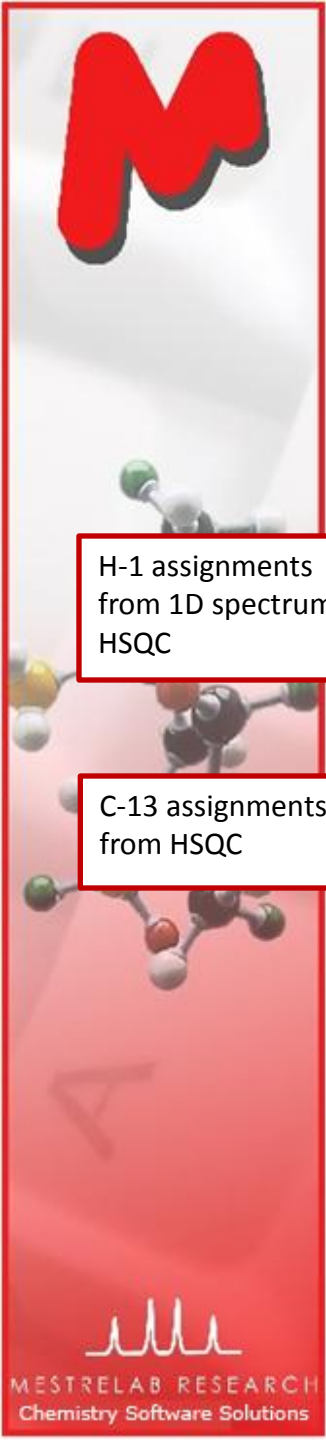
- You can first assign 1D H-1 peaks, and then assign HSQC cross peaks, or vice versa
- Assignments in one spectrum is carried over to all other spectra in the same document: All spectra in the same document are “correlated” by default
- To assign in HSQC, click **A** key to enter Assignment mode. Click on an **atom** in the structure. Next click on the cross peak to assign to it\*

H-1 assignments  
from 1D spectrum or  
HSQC

C-13 assignments  
from HSQC



\*By Default, Mnova automatically snaps to a peak top (with interpolation). Click **Shift** key one time to toggle it off if you want to manually locate the peak center.



# M

## The Assignment Table for multiple spectra

- Choose **View | Tables | Assignments** to open the Assignments Table if not yet
- The Table lists all assignment results, which can be copied to other documents
- Try **Script | Report | Assignments** to report the results in journal format

Assignments

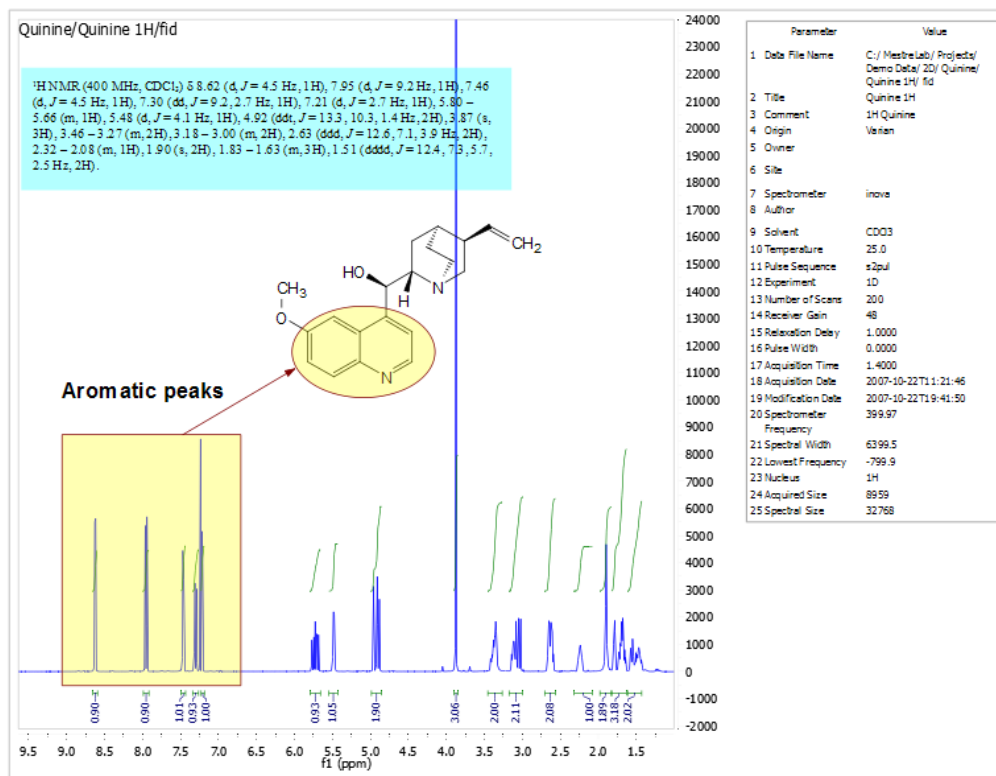
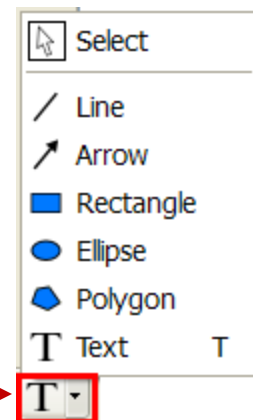
Report Copy Delete Expand Collapse Setup

Atom	Chemical SI	Predicted S	COSY	TOCSY	HSQC	HMBC	H2BC	NOESY
▲ 1 C	36.08				1', 1"	7		
H'	2.35				1			
H''	2.29				1			
2 C	79.42					7, 26		
3 C	79.40					13', 13'', ...		
4 C	133.54					57', 57'', ...		
5 C	142.27					57', 57'', ...		
▲ 6 C	72.46				6			
H	6.24				6			
▲ 7 C	75.13				7	26		
H	5.68		26		7	1, 2, 9, 28		
8 C	45.84				26			
9 C	58.92					7, 26, 21...		
10 C	203.84					11, 21', ...		
▲ 11 C	75.68				11			
H	3.82				11	10		26
▲ 12 O								
H								
▲ 13 C	27.02				13', 13'', ...			
H3	1.25				13	3		
▲ 14 C	21.95				14', 14'', ...			
H3	1.15				14			
15 O								
16 O								
17 C	81.46					24', 24'', ...		



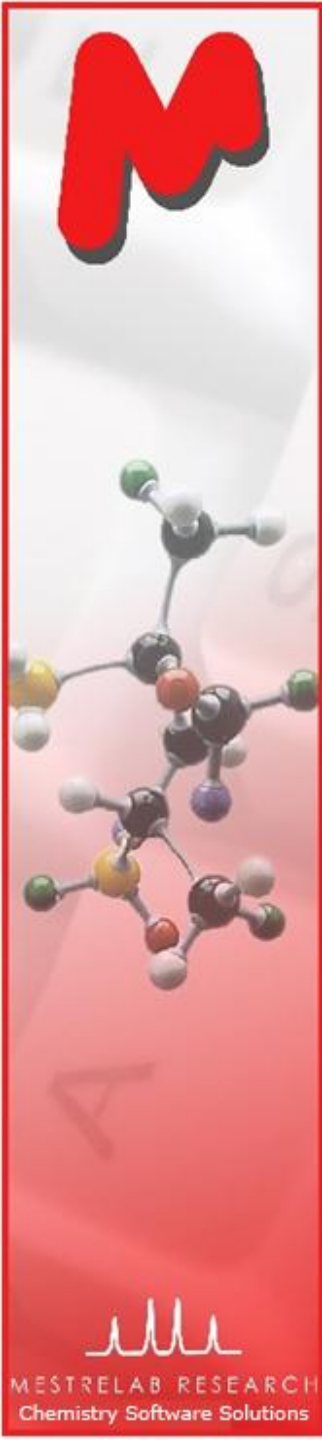
# To annotate and report manually

- Click the **Annotation Options** button at the bottom-left corner of Mnova window
- Or press **T** to insert a text box
- All objects can be customized by right clicking on it and then selecting the **Properties** command
- Tables of Peaks, Integrals, Parameters** etc can be opened by **View | Tables**. Report from there



Tips:

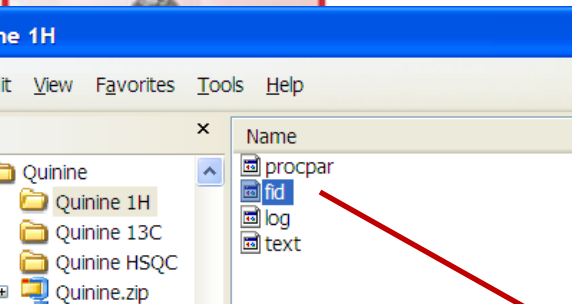
- \*Copy a **molecule** from ChemDraw or Isis/Draw, or open .mol or .sdf files
- \*Use **View | Layout Templates** menu to generate and apply layout templates, or request an **auto formatting script** from Mestrelab.
- \***Copy/paste** any object(s) to your document with high resolution
- \*Click  to export **PDF**



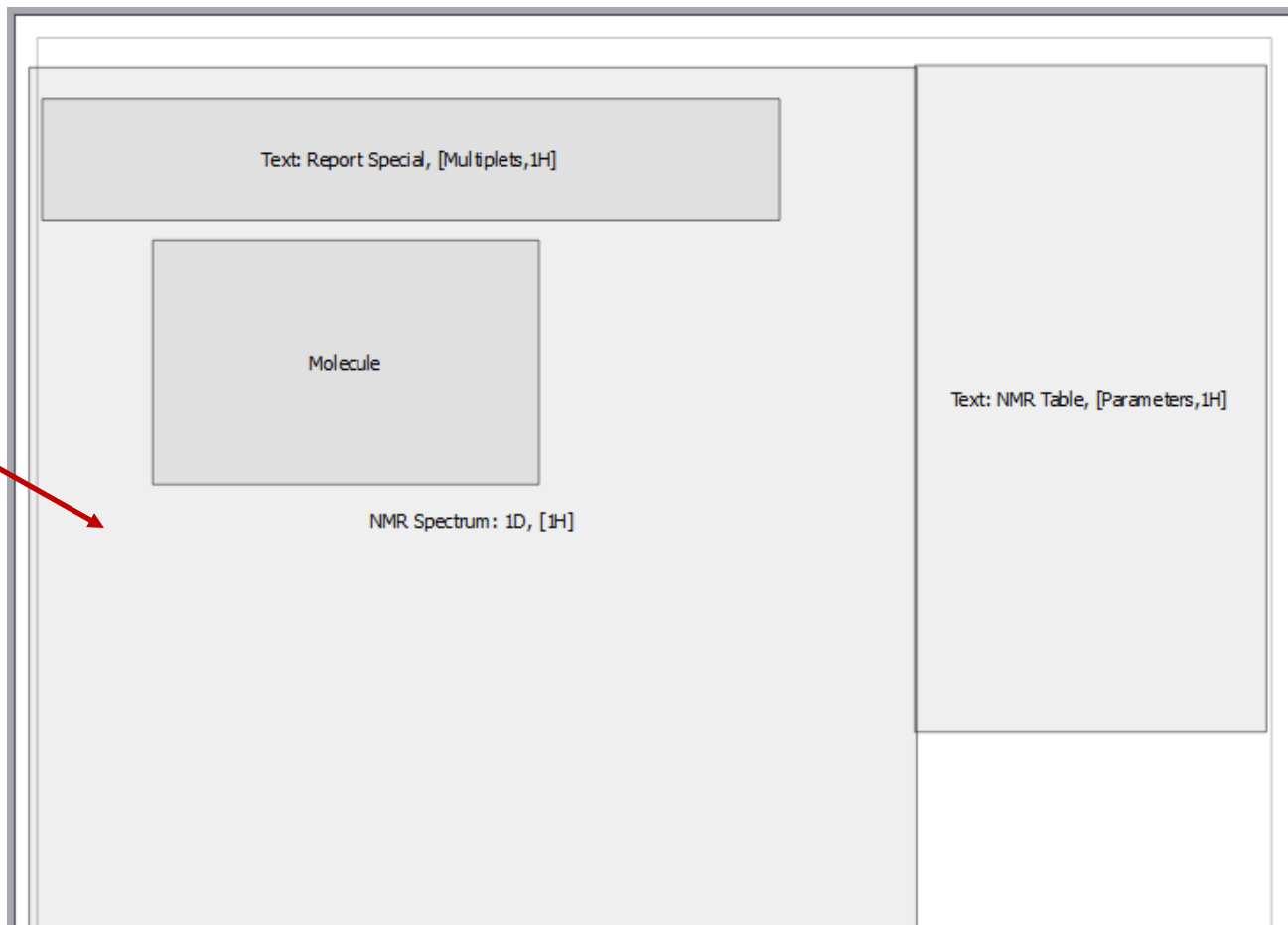


# To create layout template

- Once you are satisfied with the layout, choose **View | Layout Template | Create Layout Template Document**, and save the layout
- You can continue to edit the template
- Once ready, open a new FID or structure to the template, and they will be auto formatted to the desired size and location.
- If you have a spectrum already opened, choose **View | Layout Template | Apply Layout Template Doc** to format it

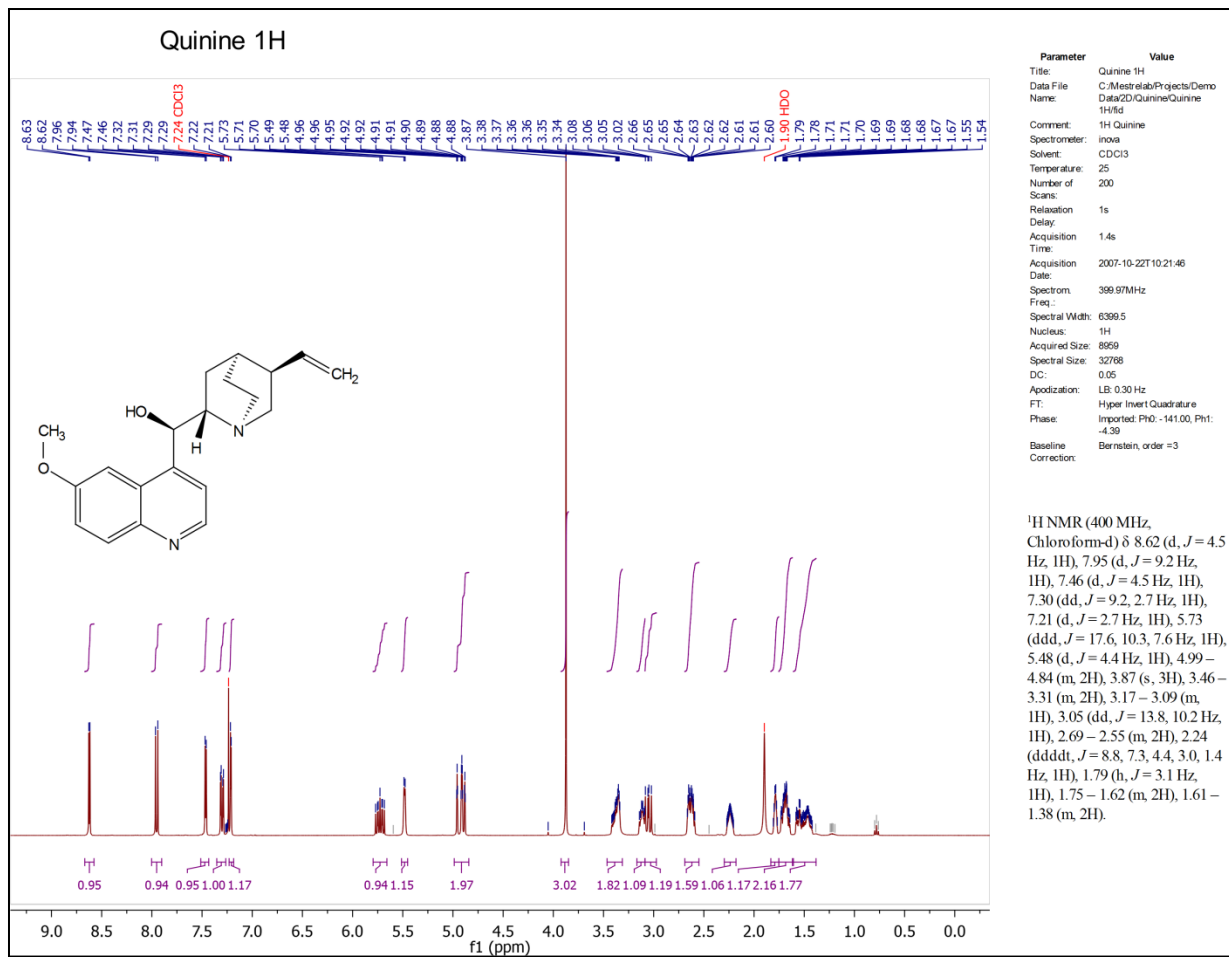


Drag & drop



# To auto format using Mnova script\*

- M Mnova has a powerful scripting engine that allows you to automate many operations, including processing, analysis and reporting
- M The following is a sample output by running a Mnova script



\* Click <http://mestrelab.com/scripts/> to download free formatting scripts. We also provide service for more complex batch processing and reporting requirements

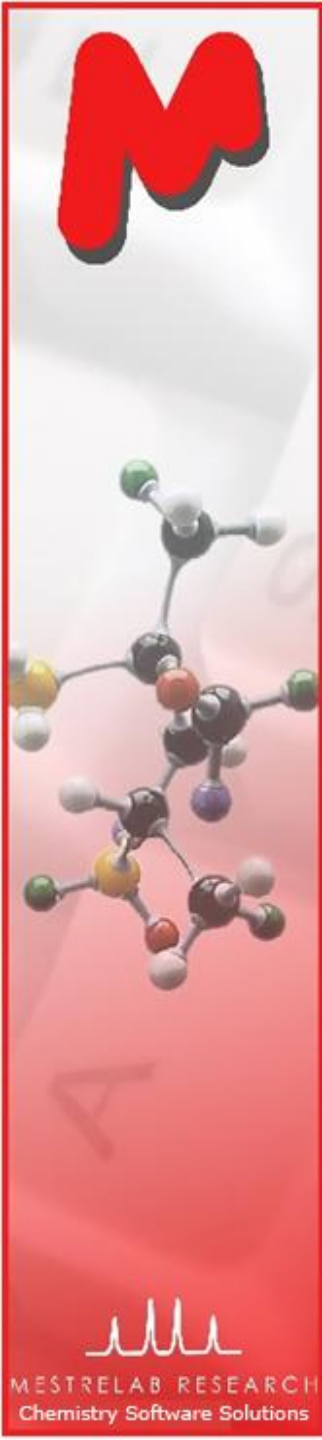
# To auto Process, Analyze and Report a 1D spectrum using an Mnova script (PAR.qs)\*

- M** You open a 1D H-1 spectrum, run this free script\* for the first time. It does the following: \*\*
  - M** Re-processing the spectrum with line broadening of 0.3 Hz, enhanced correction for Bruker Group Delay if applicable, zero-filling to double the data size or at least 64K points, and baseline correction using 3<sup>rd</sup> order Bernstein Polynomial
  - M** Automated peak picking and multiplet analysis using the current options
- M** You manually verify and correct the multiplet analysis results
- M** You run the script *again*, and it generates a report similar to the one in the previous slide
- M** You can easily customize the processing, analysis and reporting options by editing the script.
- M** This script also works for C-13 and other nucleus, in slightly different way (e.g., it picks and reports peaks instead of multiplets).
- M** This script does formatting only if it is a 2D NMR

\* Write to [chen.peng@mestrelab.com](mailto:chen.peng@mestrelab.com) and ask for PAR.qs. It's free.


To run the script, first save it on your computer. Next choose Scripts > Run Script, and open it.

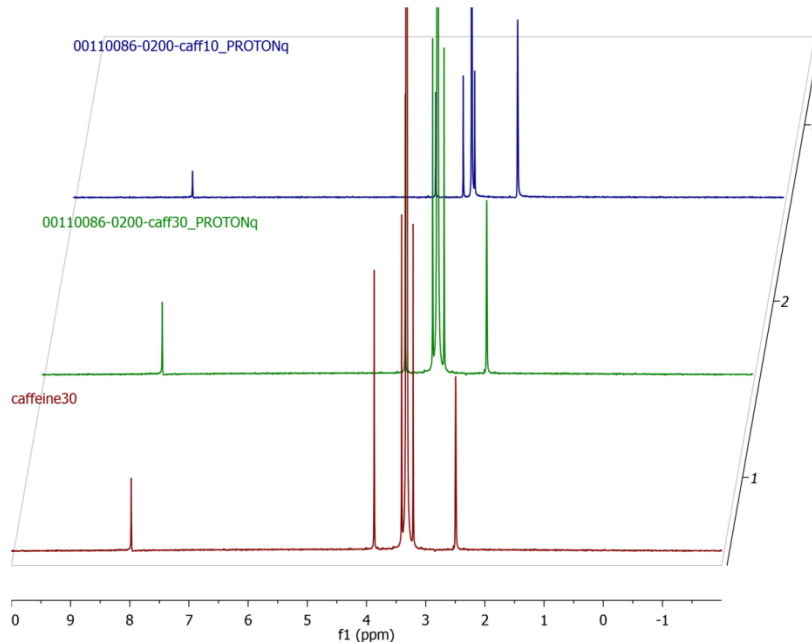
\*\* You can edit the script and customize the settings.




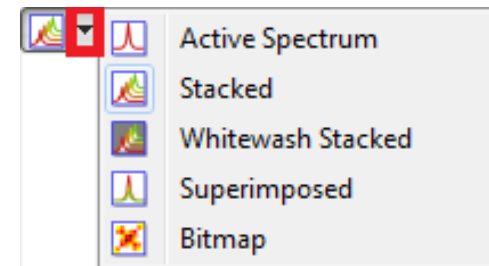
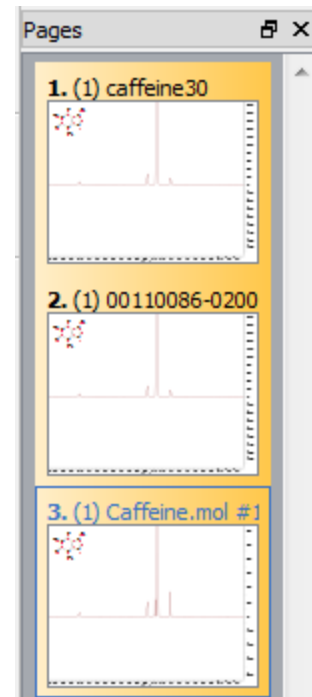



# To open and stack multiple 1D spectra

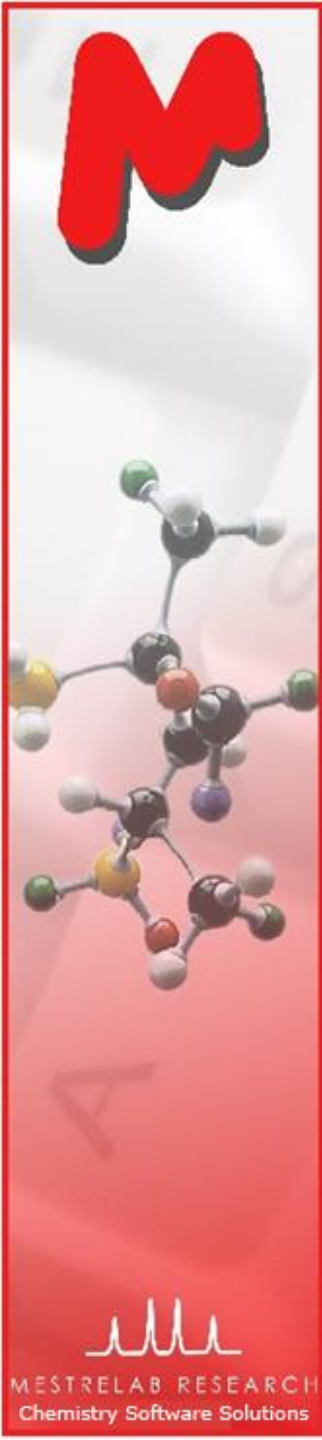
- Open several 1D spectra in the same document
- Select some or all of them in the Pages View
- Click  to stack them in a new page:



- Click  to change the display to another Stack Mode, such as the Superimposed mode

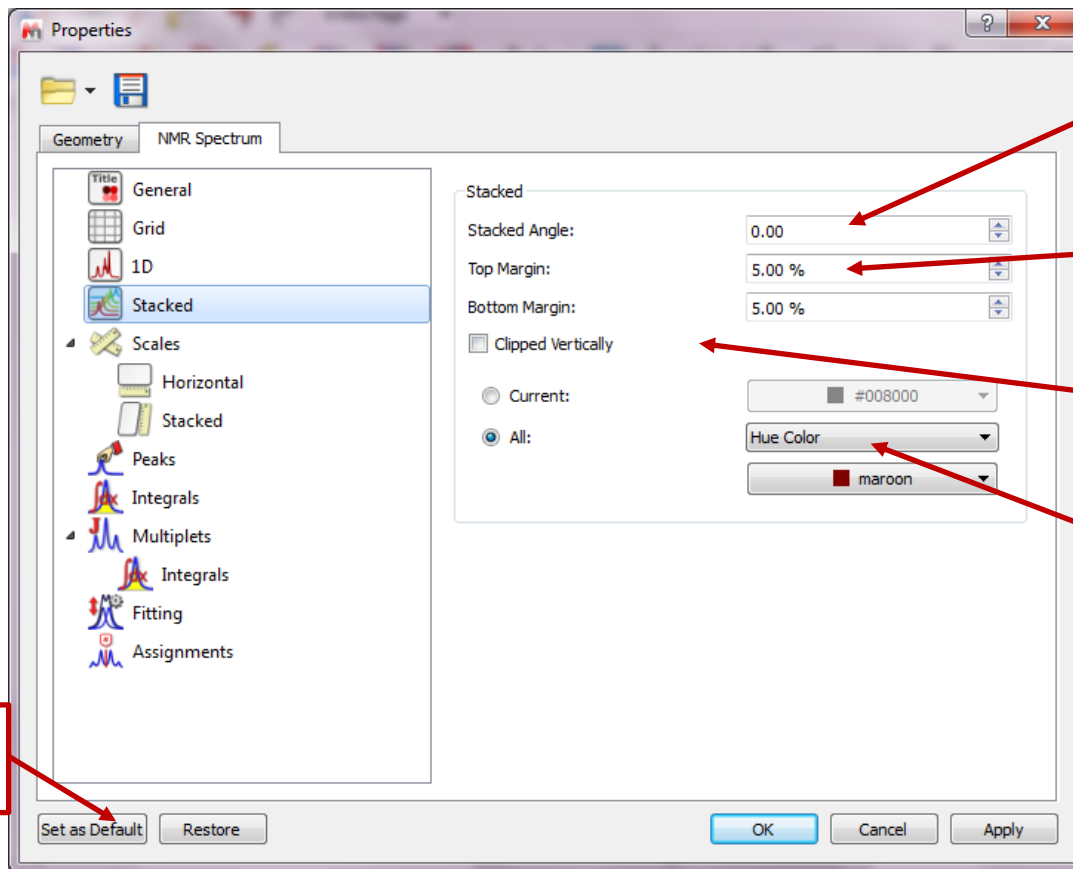


- Tip:* - You can also drag a 1D from a different page to stack it to the current page using the Pages View.
- When multiplet pages are selected, you can choose the **Superimposed** tool  to superimpose them directly.
  - If you want to stack all the 1D spectra under a certain folder, use **Scripts > Import > Directory Spectra Stack**.



# To change display properties of stacked spectra

**M** Right click on it and select **Properties**:



Enter 0 here if you don't like the tilt angle

Enlarge the top/bottom margins if you don't want to clip peaks there


Check here if you want to clip the peaks

Change colors of spectra

Click here to set the changes as default



# To handle the stacked spectra

Click  to toggle on the **Stacked Spectra Table**

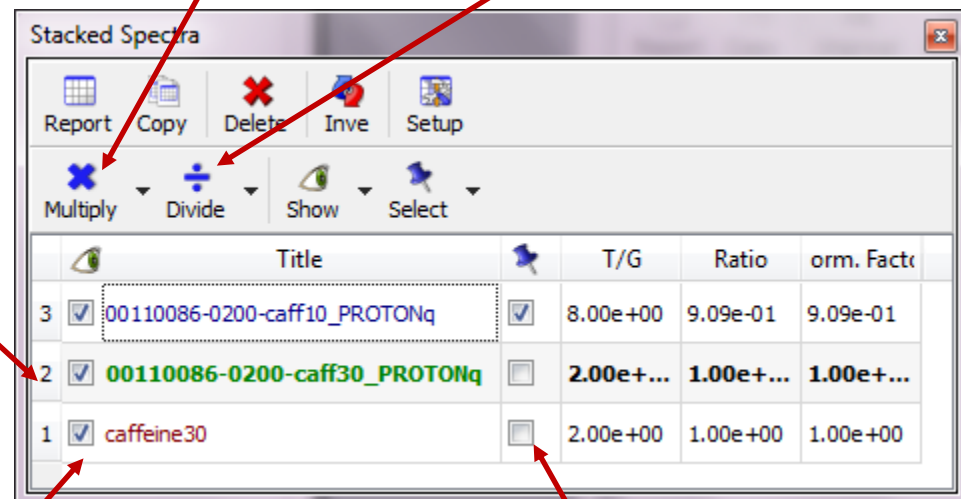
Use this table to do the following:

- Delete spectra from the stack
- Change order of the spectra in the stack
- Change the Y-intensity of selected spectra
- Choose which ones to display
- Choose which ones to adjust

To increase the Y intensity of selected or all spectra \*

To decrease Y intensity of selected or all spectra\*

Click and drag here to change the order of a spectrum in the stack



		Title		T/G	Ratio	orm. Factr
3	<input checked="" type="checkbox"/>	00110086-0200-caff10_PROTONq	<input checked="" type="checkbox"/>	8.00e+00	9.09e-01	9.09e-01
2	<input checked="" type="checkbox"/>	00110086-0200-caff30_PROTONq	<input type="checkbox"/>	2.00e+...	1.00e+...	1.00e+...
1	<input checked="" type="checkbox"/>	caffeine30	<input type="checkbox"/>	2.00e+00	1.00e+00	1.00e+00

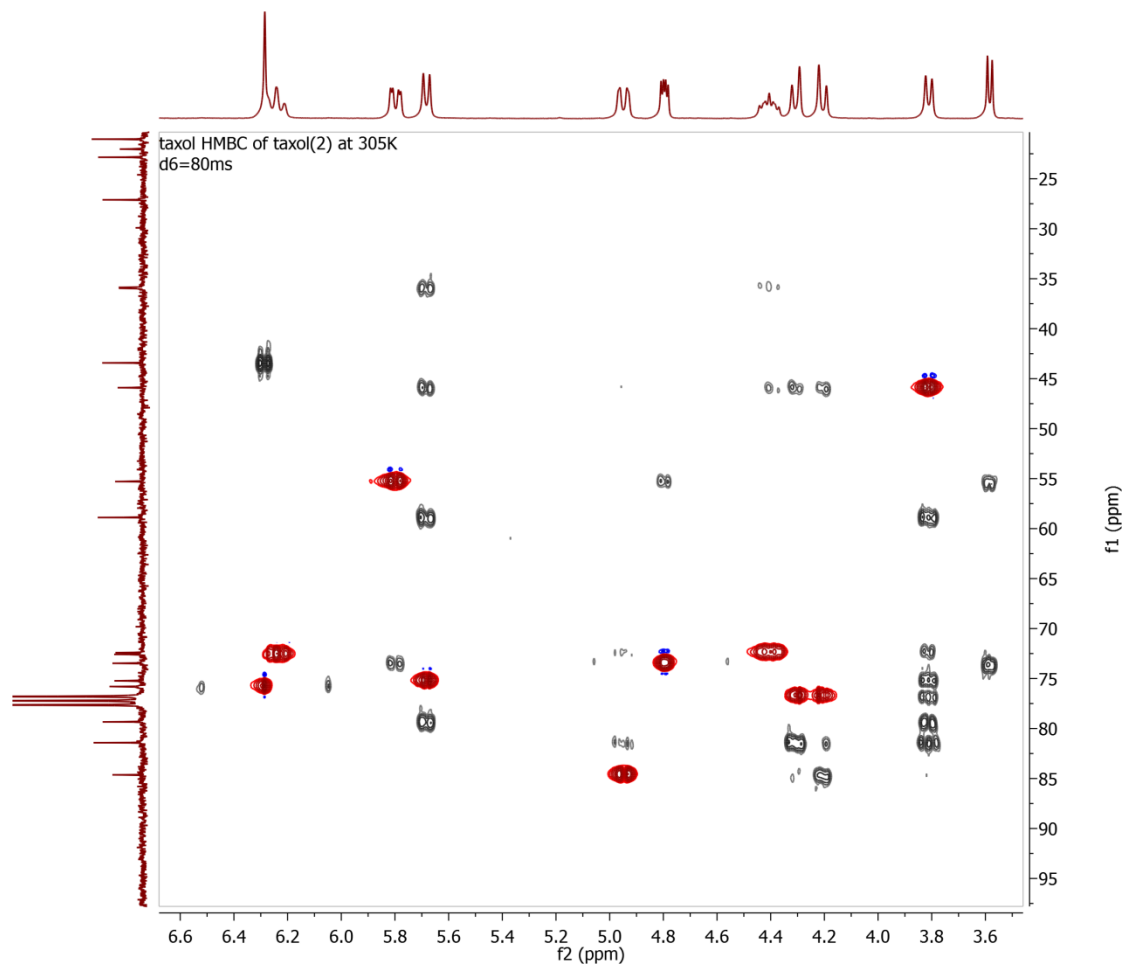
Tip: Read Help > Contents on more advanced data analysis, such as reaction monitoring, metabolomics, relaxation studies, DOSY processing etc.

Uncheck the ones you don't want to display them

Check the ones that you want to change

# To superimpose multiple 2D

- Multiple 2D can be stacked or superimposed in the same way as 1D
- Click **Shift + Up Arrow** key to change the active spectrum
- Right click on it and select **Properties** to change the color of the contours for the active spectrum





# Reviews of Mnova NMR

**A review by Tim Claridge (U. Oxford) on  
*J. Chem. Inf. Model*, 2009, 49 (4), 1136-1137:**

*"...In conclusion, I would recommend highly this software to any chemist or spectroscopist seeking a desktop package for the analysis of 1D and 2D NMR data sets. Its attractive and intuitive graphical interface combined with some smart functionality will suit less experienced users, while dedicated spectroscopists will also be impressed with the more advanced functionality it contains."*

**A review by Mark Robert Wilcott (Rice University) on  
*J. Am. Chem. Soc.*, 2009, 131 (36), 13180:**

*"...Off-line processing and the feature for generating reports are excellent. Moreover, any laboratory group with extensive NMR processing needs will find Mnova a valuable means to free up spectrometer time by moving data processing off-line. I am impressed by the concept, packaging, and utility of Mnova. Both university and industrial laboratories will find it to be an asset."*

See lists of academic and industrial customers of Mnova at <http://mestrelab.com/about-us/customers/>.





# Thank you! For more information...

- Visit [www.mestrelab.com](http://www.mestrelab.com) for free trial, manual, tutorials, prices etc
- Check **Help > Contents** in Mnova for help on specific topics
- Email to [chen.peng@mestrelab.com](mailto:chen.peng@mestrelab.com) or [support@mestrelab.com](mailto:support@mestrelab.com) for questions.

